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**UTILITY  
PATENT APPLICATION  
TRANSMITTAL**


(Only for new nonprovisional applications under 37 CFR 1.53(b))

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<p>1. <input checked="" type="checkbox"/> Fee Transmittal Form <i>(Submit an original, and a duplicate for fee processing)</i></p> <p>2. <input checked="" type="checkbox"/> Specification [Total Pages <input type="text" value="26"/> ] <i>(preferred arrangement set forth below)</i></p> <ul style="list-style-type: none"><li>- Descriptive title of the Invention</li><li>- Cross References to Related Applications</li><li>- Statement Regarding Fed sponsored R &amp; D</li><li>- Reference to Microfiche Appendix</li><li>- Background of the Invention</li><li>- Brief Summary of the Invention</li><li>- Brief Description of the Drawings <i>(if filed)</i></li><li>- Detailed Description</li><li>- Claim(s)</li><li>- Abstract of the Disclosure</li></ul> <p>3. <input checked="" type="checkbox"/> Drawing(s) (35 USC 113) [Total Sheets <input type="text" value="12"/> ]</p> <p>4. <input type="checkbox"/> Oath or Declaration [Total Pages <input type="text"/> ]</p> <ul style="list-style-type: none"><li>a. <input type="checkbox"/> Newly executed (original or copy)</li><li>b. <input type="checkbox"/> Copy from a prior application (37 CFR 1.63(d)) <i>(for continuation/divisional with Box 17 completed)</i> <i>[Note Box 5 below]</i></li><li>i. <input type="checkbox"/> DELETION OF INVENTOR(S) <small>Signed statement attached deleting inventor(s) named in the prior application, see 37 CFR 1.63(d)(2) and 1.33(b)</small></li></ul> <p>5. <input type="checkbox"/> Incorporation By Reference <i>(useable if Box 4b is checked)</i> <small>The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under Box 4b, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein.</small></p>	<p>6. <input type="checkbox"/> Microfiche Computer Program <i>(Appendix)</i></p> <p>7. <input type="checkbox"/> Nucleotide and/or Amino Acid Sequence Submission <i>(if applicable, all necessary)</i></p> <ul style="list-style-type: none"><li>a. <input type="checkbox"/> Computer Readable Copy</li><li>b. <input checked="" type="checkbox"/> Paper Copy (identical to computer copy)</li><li>c. <input checked="" type="checkbox"/> Statement verifying identity of above copies</li></ul> <p><b>ACCOMPANYING APPLICATION PARTS</b></p> <p>8. <input type="checkbox"/> Assignment Papers (cover sheet &amp; document(s))</p> <p>9. <input type="checkbox"/> 37 CFR 3.73(b) Statement <input type="checkbox"/> Power of Attorney <i>(when there is an assignee)</i></p> <p>10. <input type="checkbox"/> English Translation Document <i>(if applicable)</i></p> <p>11. <input type="checkbox"/> Information Disclosure Statement (IDS)/PTO-1449 <input type="checkbox"/> Copies of IDS Citations</p> <p>12. <input type="checkbox"/> Preliminary Amendment</p> <p>13. <input checked="" type="checkbox"/> Return Receipt Postcard (MPEP 503) <i>(Should be specifically itemized)</i></p> <p>14. <input checked="" type="checkbox"/> Small Entity <input checked="" type="checkbox"/> Statement filed in prior application, Status still proper and desired</p> <p>15. <input type="checkbox"/> Certified Copy of Priority Document(s) <i>(if foreign priority is claimed)</i></p> <p>16. <input type="checkbox"/></p> <p>17. If a <b>CONTINUING APPLICATION</b>, check appropriate box and supply the requisite information:</p> <p><input checked="" type="checkbox"/> Continuation-in-part (CIP) of prior application No: 09/346,794</p> <p><b>18. CORRESPONDENCE ADDRESS</b></p> <p>Kate H. Murashige Registration No. 29,959</p> <p>Morrison &amp; Foerster LLP 2000 Pennsylvania Avenue, N.W. Washington, D.C. 20006-1888 Telephone: (858) 720-5112 Facsimile: (202) 887-0763</p>

- ☒ If a paper is untimely filed in the above-referenced application by applicant or his/her representative, the Assistant Commissioner is hereby petitioned under 37 C.F.R. § 1.136(a) for the minimum extension of time required to make said paper timely. In the event a petition for extension of time is made under the provisions of this paragraph, the Assistant Commissioner is hereby requested to charge any fee required under 37 C.F.R. § 1.17(a)-(d) to **Deposit Account No. 03-1952**. However, the Assistant Commissioner is **NOT** authorized to charge the cost of the issue fee to the Deposit Account.

The filing fee has been calculated as follows:

FOR	NUMBER FILED	NUMBER EXTRA	RATE	CALCULATIONS
TOTAL CLAIMS	20 - 20 =	0	x \$18.00	\$0
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MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+ \$260.00	\$260.00
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TOTAL OF ABOVE CALCULATIONS =				\$1028.00
Reduction by 1/2 for filing by small entity (Note 37 C.F.R. §§ 1.9, 1.27, 1.28). If applicable, verified statement must be attached.				\$514.00
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Dated: July 6, 2000

Respectfully submitted,

By: Kate H. Murashige  
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Applicant or Patentee: SNUTCH ET AL. Attorney's Docket No. NMED.P-001-US  
 Serial or Patent No.: 09/030,482 Filed or Issued: 25 February 1998  
 For: NOVEL HUMAN CALCIUM CHANNELS AND RELATED PROBES, CELL LINES AND METHODS

**VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY STATUS**  
**(37 CFR 1.9(f) and 1.27(c)) - SMALL BUSINESS CONCERN**

I hereby declare that I am

- ☐ the owner of the small business concern identified below:  
☒ an official of the small business concern empowered to  
 act on behalf of the concern identified below:

NAME OF CONCERN NeuroMed Technologies Inc.

ADDRESS OF CONCERN 3963 W. 24<sup>th</sup> Avenue, Vancouver, Canada V6S 1M1

I hereby declare that the above identified small business concern qualifies as a small business concern as defined in 13 CFR 121.3-18, and reproduced in 37 CFR 1.9(d), for purposes of paying reduced fees under Section 41(a) and (b) of Title 35, United States Code, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third party or parties controls or has the power to control both.

I hereby declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the above-captioned invention which is described in

- ☐ the specification filed herewith  
☒ Application Serial No. 09/030,482, filed 25 February 1998  
☐ Patent No. \_\_\_\_\_, issued \_\_\_\_\_

If the rights held by the above identified small business concern are not exclusive, each individual, concern or organization having rights to the invention is listed below\* and no rights to the invention are held by any person, other than the inventor, who could not qualify as a small business concern under 37 CFR 1.9(d) or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e). \*Note: Separate verified statements are required from each named person, concern or organization having rights to the invention averring to their status as small entities. (37 CFR 1.27)

NAME \_\_\_\_\_

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☐ INDIVIDUAL ☐ SMALL BUSINESS CONCERN ☐ NONPROFIT ORGANIZATION

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I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity no longer appropriate. (37 CFR 1.28(b))

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

NAME OF PERSON SIGNING Terrance P. Snutch TITLE President

ADDRESS OF PERSON SIGNING 3963 West 24<sup>th</sup> Ave, Vancouver BC V6S 1M1

SIGNATURE T.P. Snutch DATE May 27, 1997

## MAMMALIAN T-TYPE CALCIUM CHANNELS

This application is a continuation-in-part of application No. 09/346,794 filed  
2 July 1999 which is a continuation-in-part of application No. 09/030,428 filed  
25 February 1998 which claims priority from Provisional Application No. 60/039,204  
5 filed 28 February 1997. The disclosures of these applications are incorporated by  
reference herein.

### TECHNICAL FIELD

The invention relates to T-type calcium channel encoding sequences, expression  
of these sequences, and methods to screen for compounds which antagonize calcium  
10 channel activity. The invention is also related to molecular tools derived from knowledge  
of the molecular structure of T-type calcium channels.

### BACKGROUND OF THE INVENTION

The rapid entry of calcium into cells is mediated by a class of proteins called  
voltage-gated calcium channels. Calcium channels are a heterogeneous class of  
15 molecules that respond to depolarization by opening a calcium-selective pore through the  
plasma membrane. The entry of calcium into cells mediates a wide variety of cellular  
and physiological responses including excitation-contraction coupling, hormone secretion  
and gene expression. In neurons, calcium entry directly affects membrane potential and  
contributes to electrical properties such as excitability, repetitive firing patterns and  
20 pacemaker activity. Miller, R.J. (1987) "Multiple calcium channels and neuronal  
function." *Science* 235:46-52. Calcium entry further affects neuronal functions by  
directly regulating calcium-dependent ion channels and modulating the activity of  
calcium-dependent enzymes such as protein kinase C and calmodulin-dependent protein  
kinase II. An increase in calcium concentration at the presynaptic nerve terminal triggers  
25 the release of neurotransmitter. Calcium entry also plays a role in neurite outgrowth and  
growth cone migration in developing neurons and has been implicated in long-term  
changes in neuronal activity.

In addition to the variety of normal physiological functions mediated by calcium channels, they are also implicated in a number of human disorders. Recently, mutations identified in human and mouse calcium channel genes have been found to account for several disorders including, familial hemiplegic migraine, episodic ataxia type 2, cerebellar ataxia, absence epilepsy and seizures. Fletcher, *et al.* (1996) "Absence epilepsy in tottering mutant mice is associated with calcium channel defects." *Cell* 87:607-617; Burgess, *et al.* (1997) "Mutation of the Ca<sup>2+</sup> channel  $\beta$  subunit gene Cchb4 is associated with ataxia and seizures in the lethargic (lh) mouse." *Cell* 88:385-392; Ophoff, *et al.* (1996) "Familial hemiplegic migraine and episodic ataxia type-2 are caused by mutations in the Ca<sup>2+</sup> channel gene CACNL1A4." *Cell* 87:543-552; Zhuchenko, O., *et al.* (1997) "Autosomal dominant cerebellar ataxia (SCA6) associated with the small polyglutamine expansions in the  $\alpha$ 1A-voltage-dependent calcium channel." *Nature Genetics* 15:62-69.

The clinical treatment of some disorders has been aided by the development of therapeutic calcium channel antagonists. Janis, *et al.* (1991) in *Calcium Channels: Their Properties, Functions, Regulation and Clinical Relevance*. CRC Press, London.

Native calcium channels have been classified by their electrophysiological and pharmacological properties as T, L, N, P and Q types (for reviews see McCleskey, *et al.* (1991) "Functional properties of voltage-dependent calcium channels." *Curr. Topics Membr.* 39: 295-326, and Dunlap, *et al.* (1995) "Exocytotic Ca<sup>2+</sup> channels in mammalian central neurons." *Trends Neurosci.* 18:89-98.). T-type (or low voltage-activated) channels describe a broad class of molecules that activate at negative potentials and are highly sensitive to changes in resting potential. The L, N, P and Q-type channels activate at more positive potentials and display diverse kinetics and voltage-dependent properties. There is some overlap in biophysical properties of the high voltage-activated channels, consequently pharmacological profiles are useful to further distinguish them. L-type channels are sensitive to dihydropyridine (DHP) agonists and antagonists, N-type channels are blocked by the *Conus geographus* peptide toxin,  $\omega$ -conotoxin GVIA, and P-type channels are blocked by the peptide  $\omega$ -agatoxin IVA from the venom of the funnel web spider, *Agelenopsis aperta*. A fourth type of high voltage-activated Ca channel (Q-type) has been described, although whether the Q- and P-type channels are distinct molecular entities is controversial (Sather *et al.* (1993) "Distinctive biophysical and

pharmacological properties of class A (B1) calcium channel  $\alpha_1$  subunits.” *Neuron* 11:291-303; Stea, *et al.* (1994) “Localization and functional properties of a rat brain  $\alpha_1A$  calcium channel reflect similarities to neuronal Q- and P-type channels.” *Proc Natl Acad Sci (USA)* 91:10576-10580; Bourinet, E., *et al.* (1999) *Nature Neuroscience* 2:407-415).

5 Several types of calcium conductances do not fall neatly into any of the above categories and there is variability of properties even within a category suggesting that additional calcium channels subtypes remain to be classified.

Biochemical analyses show that neuronal high-threshold calcium channels are heterooligomeric complexes consisting of three distinct subunits ( $\alpha_1$ ,  $\alpha_2\delta$  and  $\beta$ )

10 (reviewed by De Waard, *et al.* (1997) in *Ion Channels*, Volume 4, edited by Narahashi, T. Plenum Press, New York). The  $\alpha_1$  subunit is the major pore-forming subunit and contains the voltage sensor and binding sites for calcium channel antagonists. The mainly extracellular Alternatively, the  $\alpha_2$  subunit is disulphide-linked to the transmembrane  $\delta$  subunit and both are derived from the same gene and are proteolytically

15 cleaved *in vivo*. The  $\beta$  subunit is a non-glycosylated, hydrophilic protein with a high affinity of binding to a cytoplasmic region of the  $\alpha_1$  subunit. A fourth subunit,  $\gamma$ , is unique to L-type Ca channels expressed in skeletal muscle T-tubules. The isolation and characterization of  $\gamma$ -subunit-encoding cDNAs is described in U.S. Patent No. 5,386,025 which is incorporated herein by reference.

Molecular cloning has revealed the cDNA and corresponding amino acid sequences of six different types of  $\alpha_1$  subunits ( $\alpha_{1A}$ ,  $\alpha_{1B}$ ,  $\alpha_{1C}$ ,  $\alpha_{1D}$ ,  $\alpha_{1E}$  and  $\alpha_{1S}$ ) and four types of  $\beta$  subunits ( $\beta_1$ ,  $\beta_2$ ,  $\beta_3$  and  $\beta_4$ ) (reviewed in Stea, A., Soong, T.W. and Snutch, T.P. (1994) “Voltage-gated calcium channels.” in *Handbook of Receptors and Channels*. Edited by R.A. North, CRC Press). A comparison of the amino acid

20 sequences of these  $\alpha_1$  subunits is included in this publication, which is incorporated herein by reference. PCT Patent Publication WO 95/04144, which is incorporated herein by reference, discloses the sequence and expression of  $\alpha_{1E}$  calcium channel subunits.

As described in Stea, A., *et al.* (1994) (*supra*), the  $\alpha_1$  subunits are generally of the order of 2000 amino acids in length, ranging from 1873 amino acids in  $\alpha_{1S}$  derived from

30 rabbit to 2424 amino acids in  $\alpha_{1A}$  derived from rabbit. Generally, these subunits contain 4 internal homologous repeats (I-IV) each having six putative alpha helical membrane

spanning segments (S1-S6) with one segment (S4) having positively charged residues every 3rd or 4th amino acid. There are a minority of a splice variant exceptions. Between domains II and III there is a cytoplasmic domain which is believed to mediate excitation-contraction coupling in  $\alpha_{1S}$  and which ranges from 100-400 amino acid residues among the subtypes. The domains I-IV make up roughly 2/3 of the molecule and the carboxy terminus adjacent to the S6 region of domain IV is believed to be on the intracellular side of the calcium channel. There is a consensus motif (QQ-E-L-GY-WI-E) in all of the subunits cloned and described in Stea, A., *et al.* (supra) downstream from the domain I S6 transmembrane segment that is a binding site for the  $\beta$  subunit.

PCT publication WO 98/38301, which describes the work of the inventors herein, and which is incorporated herein by reference, reports the first description of the molecular composition of T-type calcium channel  $\alpha_1$  subunits. The present application describes full-length genes for 3 mammalian subtypes,  $\alpha_{1G}$ ,  $\alpha_{1H}$ , and  $\alpha_{1I}$  associated with T-type calcium channels.

In some expression systems the high threshold  $\alpha_1$  subunits alone can form functional calcium channels although their electrophysiological and pharmacological properties can be differentially modulated by coexpression with any of the four  $\beta$  subunits. Until recently, the reported modulatory affects of  $\beta$  subunit coexpression were to mainly alter kinetic and voltage- dependent properties. More recently it has been shown that  $\beta$  subunits also play crucial roles in modulating channel activity by protein kinase A, protein kinase C and direct G-protein interaction. (Bourinet, *et al.* (1994) "Voltage-dependent facilitation of a neuronal  $\alpha_{1C}$  L-type calcium channel." *EMBO J.* 13: 5032-5039; Stea, *et al.* (1995) "Determinants of PKC- dependent modulation of a family of neuronal calcium channels." *Neuron* 15:929-940; Bourinet, *et al.* (1996) "Determinants of the G-protein-dependent opioid modulation of neuronal calcium channels." *Proc. Natl. Acad. Sci. (USA)* 93: 1486-1491.)

Because of the importance of calcium channels in cellular metabolism and human disease, it would be desirable to identify the remaining classes of  $\alpha_1$  subunits, and to develop expression systems for these subunits which would permit the study and characterization of these calcium channels, including the study of pharmacological modulators of calcium channel function.

## DISCLOSURE OF THE INVENTION

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The present invention provides sequences for a novel mammalian calcium channel subunits of T-type calcium channels, which we have labeled as  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits. Knowledge of the sequences of these calcium channel subunits may be used in the development of probes for mapping the distribution and expression of the subunits in target tissues. In addition, as the molecular structure of the  $\alpha_1$  subunits of these T-type calcium channels has been elucidated, it is possible to identify those portions which reside extracellularly and thus to design peptides to elicit antibodies which can be employed to assess the location and level of expression of T-type calcium channels. In addition, these subunits, either alone or assembled with other proteins, can produce functional calcium channels, which can be evaluated in model cell lines to determine the properties of the channels containing the subunits of the invention. These cell lines can be used to evaluate the effects of pharmaceuticals and/or toxic substances on calcium channels incorporating  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits. The resulting identified compounds are useful in treating conditions where undesirable T-type calcium channel activity is present. These conditions include epilepsy, sleep disorders, mood disorders, cardiac hypertrophy and arrhythmia and hypertension, among others. In addition, antisense and triplex nucleotide sequences can be designed to inhibit the production of T-type calcium channels.

In a preferred embodiment the  $\alpha_1$  subunits are other than those encoded by SEQ. ID. NO: 17; in another preferred embodiment the  $\alpha_1$  subunits are other than those encoded by sequences that include SEQ. ID. NO: 19 and SEQ. ID. NO: 21. In another preferred embodiment, probes representing portions of or all of SEQ. ID. NOS. 1-22 or 13-21 are excluded.

## BRIEF DESCRIPTION OF THE DRAWINGS

Figs. 1A and B show a comparison of the waveforms and current voltage relationship for  $\alpha_{1G}$ ;

Figs. 2A and B show a comparison of the waveforms and current voltage relationship for  $\alpha_{1I}$  calcium channels.



Fig. 3 shows a comparison of the steady state inactivation profiles of the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channels.

Figs. 4A-C show a comparison of the inactivation kinetics of the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channels.

5            Figures 5A and 5B show the construction of the human  $\alpha_{1G}$  cDNA complete sequence from partial clones.

Figure 6 shows the nucleotide and deduced amino acid sequence of human T-type calcium channel  $\alpha_{1G}$ .

10            Figure 7 shows a comparison of the waveforms and current voltage relationship for human  $\alpha_{1G}$  calcium channel.

Figure 8 shows the characteristic pore pattern for T-type channels.

#### MODES OF CARRYING OUT THE INVENTION

The present invention includes the following aspects for which protection is sought:

15            (a)    novel mammalian (including human) calcium channel subunits and DNA sequences encoding such subunits. Specifically, the invention encompasses an at least partially purified DNA molecule comprising a sequence of nucleotides that encodes an  $\alpha_1$  subunit of a T-type calcium channel, and such  $\alpha$  subunits *per se*. It will be appreciated that polymorphic variations may be made or may exist in the DNA of some individuals  
20            leading to minor deviations in the DNA or amino acids sequences from those shown which do not lead to any substantial alteration in the function of the calcium channel. Such variations, including variations which lead to substitutions of amino acids having similar properties are considered to be within the scope of the present invention. Thus, in one embodiment, the present application claims DNA molecules which encode  $\alpha_1$   
25            subunits of mammalian T-type calcium channels, and which hybridize under conditions of medium (or higher) hybridization stringency with one or another of the specific sequences disclosed in this application. This level of hybridization stringency is generally sufficient given the length of the sequences involved to permit recovery of the subunits within the scope of the invention from mammalian DNA libraries.

Alternatively, the T-type calcium channels of the invention are recognized by their functional characteristic of low voltage gating along with defined structural characteristics which classify them as  $\alpha_1$  calcium channel subunits and also characterize them as of the T-type. By virtue of the present invention, these characteristics have been elucidated as follows:

One distinguishing feature of the  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  T-type channels over other types of calcium channels and sodium channels is that the pore region (P-region) in each of the four structural domains contains a diagnostic amino acid sequence implicated in channel permeability. Figure 8 shows that the T-type channels contain the residues glutamate/glutamate/aspartate/aspartate (single letter amino acid code: EEDD) in the P-regions of domains I-IV. In contrast, figure 8 shows that in sodium (Na) channels the P-region of the four domains contains the residues: aspartate/glutamate/lysine/alanine (single letter amino acid code: DEKA), while high threshold calcium channels such as the L-type channel contain the residues: glutamate/glutamate/glutamate/glutamate (single letter amino acid code: EEEE). The  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  T-type channels are also distinct in this region compared to other types of ion channels including the *C. elegans* C11D2.6 and C27F2.3 and the rat NIC-channel (Figure 8).

A second distinguishing characteristic of the  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  T-type channels compared to other types of calcium channels is that they do not contain a  $\beta$  subunit binding consensus sequence in the cytoplasmic linker separating domains I and II. In contrast, all high threshold calcium channels contain a consensus sequence (single letter amino acid code: QQ-E--L-GY--WI---E) shown to physically interact with the calcium channel  $\beta$  subunit (Pragnell, M., De Waard, M., Mori, Y., Tanabe, T., Snutch, T.P. & Campbell, K.P., 1994, Nature 368:67-70). Thus it appears the presence of a  $\beta$  subunit does not modify activity, nor is its presence required.

A third distinguishing characteristic of the ( $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  T-type channels is that they do not possess an EF-hand calcium binding motif in the region carboxyl to domain IV S6. In contrast, all high threshold calcium channels contain a consensus sequence that is closely related to the EF-hand domain found in certain calcium binding

proteins (de Leon, M., Wang, Y., Jones, L., Perez-Reyes, E., Wei, X., Soong, T.W., Snutch, T.P. & Yue, D.T., 1995, Science270: 1502-1506).

Thus, as defined herein, "T-type calcium channel  $\alpha_1$  subunits" refers to subunits which contain these structural characteristics.

5 Alternatively, the T-type  $\alpha_1$  subunit molecules can be defined by homology to the human and rat nucleotide and amino acid sequences described herein. Thus, T-type  $\alpha_1$  subunits will typically have at least 50%, preferably 70% homology in terms of amino acid sequence or encoding nucleotide sequence to the sequences set forth in SEQ ID NOS. 23-28 herein or those shown in Figure 6. Preferably, the homology will be at least  
10 80%, more preferably 90%, and most preferably 95%, 97%, 98% or 99%.

Relative homology may also be defined in terms of specific regions; as set forth above, certain regions of T-type channel  $\alpha_1$  subunits have very high homologies while other regions, such as the cytoplasmic region between domains II and III have less homology. Thus, T-type  $\alpha_1$  subunits will have over 75% homology; preferably over 85%  
15 or over 95% homology, more preferably over 98% homology in domains I-IV to those of SEQ. ID. NOS. 23-28 or Figure 6. The degree of homology in the cytoplasmic region between domains II and III may be substantially less, *e.g.*, only 25% homology, preferably, 50% homology or more preferably 60% homology. Similarly, the intracellular region downstream of domain IV may be less homologous than within  
20 domains I-IV.

(b) polynucleotide sequences useful as probes in screening human cDNA libraries for genes encoding these novel calcium channel subunits. These probes can also be used in histological assay to determine the tissue distribution of the novel calcium channel subunits.

25 As set forth above, the elucidation herein of the structural features of T-type subunits permits the selection of appropriate probes by selecting portions of the encoding nucleotide sequence that are particularly characteristic of this type. As set forth above, for example, T-type subunits have particular patterns of amino acids in the pore forming units as set forth in Figure 8. Alternatively, multiple probes might be used to distinguish  
30 other subunits, such as probes which represent the  $\beta$ -binding domain missing from the T-

type  $\alpha_1$  subunits combined with a probe representing a consensus sequence for calcium channel  $\alpha$  subunits in general.

(c) at least partially purified  $\alpha_1$  subunits and related peptides for mammalian T-type calcium channels. These proteins and peptides can be used to generate polyclonal or monoclonal antibodies to determine the cellular and subcellular distribution of T-type calcium channel subunits.

Again, by virtue of the elucidation of the amino acid sequence of T-type  $\alpha_1$  subunits, it is well within the ordinary skill in the art to determine which regions of the channel are displayed extracellularly and to select these regions for the generation of antibodies.

(d) eukaryotic cell lines expressing the novel calcium channel subunits. These cell lines can be used to evaluate compounds as pharmacological modifiers of the function of the novel calcium channel subunits.

(e) a method for evaluating compounds as pharmacological modifiers of the function of the novel calcium channel subunits using the cell lines expressing those subunits alone or in combination with other calcium channel subunits.

(f) Use of the compounds identified as set forth above for the treatment of conditions which are associated with undesired calcium channel activity.

These diseases include, but are not limited to; epilepsy, migraine, ataxia, schizophrenia, hypertension, arrhythmia, angina, depression, and Parkinson's disease; characterization of such associations and ultimately diagnosis of associated diseases can be carried out with probes which bind to the wild-type or defective forms of the novel calcium channels.

T-type channels in particular are responsible for rebound burst firing in central neurons and are implicated in normal brain functions such as slow-wave sleep and in neurological disorders such as epilepsy and mood disorders. They are also important in pacemaker activity in the heart, hormone secretion and fertilization, and are associated with disease states such as cardiac hypertrophy and hypertension.

As used in the specification and claims of this application, the term "T-type calcium channel" refers to a voltage-gated calcium channel having a low activation voltage, generally less than -50 mV, and most commonly less than -60 mV. T-type

calcium channels also exhibit comparatively negative steady-state inactivation properties and slow deactivation kinetics. The terms " $\alpha_1$  subunit" or " $\alpha_1$  calcium channel" refer to a protein subunit of a calcium channel which is responsible for pore formation and contains the voltage sensor and binding sites for calcium channel agonists and antagonists. Such subunits may be independently functional as calcium channels or may require the presence of other subunit types for complete functionality.

As used in the specification and claims of this application, the phrase "at least partially purified" refers to DNA or protein preparations in the which the specified molecule has been separated from adjacent cellular components and molecules with which it occurs in the natural state, either by virtue of performing a physical separation process or by virtue of making the DNA or protein molecule in a non-natural environment in the first place. The term encompasses cDNA molecules and expression vectors.

In accordance with the present invention, we have identified mammalian DNA sequences which code for novel T-type calcium channel  $\alpha_1$  subunits. These subunits are believed to represent new types of  $\alpha_1$  subunits of mammalian voltage-dependent calcium channels which have been designated as types  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$ .

A Bacterial Artificial Chromosome (BAC) sequence (bK206c7) was identified from sequences in Sanger Genome Sequencing Center (Cambridge, U.K.) and the Washington University Genome Sequencing Center (St. Louis, MO) that contains a nucleotide sequence encoding the  $\alpha_{1I}$  subunit of human T-type calcium channel. The rationale for this identification is set forth in WO 98/38301, incorporated herein by reference. The relevant nucleotide sequence and the translated amino acid sequence containing 1854 amino acids are set forth in SEQ ID NOS:17 and 18.

As described in WO 98/38031, using PCR cloning techniques to identify relevant sequences within a human brain total RNA preparation, we confirmed that the novel  $\alpha_{1I}$  calcium channel subunit is present in human brain. Subcloning of the 567 nt PCR product (SEQ. ID NO. 19, amino acids SEQ. ID NO. 20) and subsequent sequencing thereof showed that this product corresponds to the derived sequence from the bK206c7 BAC genomic sequence, the nucleotide sequence of which is given as SEQ ID NO. 17 (amino acid sequence SEQ. ID NO.18). The same experiment was performed using a rat

brain RNA preparation and resulted in recovery of a substantially identical PCR product. (SEQ ID. NO. 21). The protein encoded by the rat PCR product (SEQ ID NO. 22) is 96% identical to the human PCR product (SEQ. ID NO. 20).

These sequences, which encode a partial subunit were used as a basis for constructing full length human or rat  $\alpha_{1I}$  clones. Briefly, the subcloned  $\alpha_{1I}$  PCR product is radiolabeled by random hexamer priming according to standard methods (See, Sambrook, J., Fritsch, E.F. and Maniatis, T. (1989) *Molecular Cloning, A Laboratory Manual*. Cold Spring Harbor Press) and used to screen commercial human brain cDNA libraries (Stratagene, La Jolla, CA). The screening of cDNA libraries follows standard methods and includes such protocols as infecting bacteria with recombinant lambda phage, immobilizing lambda DNA to nitrocellulose filters and screening under medium hybridization stringency conditions with radiolabeled probe. cDNA clones homologous to the probe are identified by autoradiography. Positive clones are purified by sequential rounds of screening.

Following this protocol, most purified cDNA's are likely to be partial sequence clones due to the nature of the cDNA library synthesis. Full length clones are constructed from cDNA's which overlap in DNA sequence. Restriction enzyme sites which overlap between cDNAs are used to ligate the individual cDNA's to generate a full-length cDNA. For subsequent heterologous expression, the full-length cDNA is subcloned directly into an appropriate vertebrate expression vector, such as pcDNA-3 (Invitrogen, San Diego, CA) in which expression of the cDNA is under the control of a promoter such as the CMV major intermediate early promoter/enhancer. Other suitable expression vectors include, for example, pMT2, pRC/CMV, pcDNA3.1 and pCEP4.

Following these protocols, full length mammalian  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  calcium channel subunit cDNAs were isolated by using the 567 base pair human fragment (SEQ. ID NO. 19) to screen a rat brain cDNA library. Sequencing of the recovered sequences identified the three distinct classes of calcium channel subunits which have been denominated herein as  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits. For each class of subunit, complete sequencing of the largest cDNA confirmed that it represented only a portion of the predicted calcium channel coding region. Complete sequences for the three new subunits were obtained by rescreening the rat brain cDNA library with probes derived from the

partial length cDNAs to obtain overlapping segments. These segments were combined to form a complete gene by restriction digestion and ligation. The complete cDNA sequences of the rat  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits are given by SEQ. ID NOS. 23, 25 and 27, respectively. Corresponding amino acid sequences are given by SEQ. ID NOS. 24, 26 and 28. The same techniques are employed to recover human sequences by screening of a human or other mammalian library. Thus, for example, partial length human sequences for  $\alpha_{1G}$  and  $\alpha_{1H}$  T- type calcium channels have been recovered using the same probe (SEQ. ID NO. 19) and the full length rat  $\alpha_{1I}$  cDNA (SEQ. ID. NO. 27) has been used to recover a partial length DNA encoding a human  $\alpha_{1I}$  T-type calcium channel. The DNA and amino acid sequences for these partial length human calcium channels are given by SEQ. ID NOS. 30-35. A complete coding sequence for human  $\alpha_{1G}$  was also obtained and is set forth, along with the deduced amino acid reference, in Figure 6.

Once the full length cDNA is cloned into an expression vector, the vector is then transfected into a host cell for expression. Suitable host cells include *Xenopus* oocytes or mammalian cells such as human embryonic kidney cells as described in International Patent Publication No. WO 96/39512 which is incorporated herein by reference and Ltk cells as described in US Patent No. 5,386,025 which is incorporated herein by reference. Transfection into host cells may be accomplished by microinjection, lipofection, glycerol shock, electroporation calcium phosphate or particle-mediated gene transfer. The vector may also be transfected into host cells to provide coexpression of the novel  $\alpha_1$  subunits with other  $\alpha$  subunits, such as an  $\alpha_2\delta$  subunit or  $\gamma$  subunit.

To confirm that the three full length cDNAs (SEQ. ID NOS. 23, 25 and 27) encoded functional calcium channels, the  $\alpha_{1G}$  and  $\alpha_{1I}$  cDNAs were transiently transfected into human embryonic kidney cells and evaluated using electrophysiological recording techniques. The results are consistent with a role of these subunits in native T-type channels in nerve, muscle and endocrine cells. Similarly, a full length clone encoding human  $\alpha_{1G}$  T-type subunit was recovered and verified to have the characteristic properties of T-type channels.

The resulting cell lines expressing functional calcium channels including the novel  $\alpha_1$  subunits of the invention can be used test compounds for pharmacological activity with respect to these calcium channels. Thus, the cell lines are useful for

screening compounds for pharmaceutical utility. Such screening can be carried out using several available methods for evaluation of the interaction, if any, between the test compound and the calcium channel. One such method involves the binding of radiolabeled agents that interact with the calcium channel and subsequent analysis of equilibrium binding measurements including but not limited to, on rates, off rates,  $K_d$  values and competitive binding by other molecules. Another such method involves the screening for the effects of compounds by electrophysiological assay whereby individual cells are impaled with a microelectrode and currents through the calcium channel are recorded before and after application of the compound of interest. Another method, high-throughput spectrophotometric assay, utilizes the loading the cell lines with a fluorescent dye sensitive to intracellular calcium concentration and subsequent examination of the effects of compounds on the ability of depolarization by potassium chloride or other means to alter intracellular calcium levels. Compounds to be tested as agonists or antagonists of the novel  $\alpha_{1I}$  calcium channel subunits are combined with cells that are stably or transiently transformed with a DNA sequence encoding the  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  calcium channel subunits of the invention and monitored using one of these techniques.

Compounds which are shown to modulate the activity of calcium channels can then be used in pharmaceutical compositions for the treatment associated with inappropriate T-type calcium channel activity. Such conditions may also include those with inappropriate calcium channel activity in general since such activity may be modified by enhancing or decreasing T-type channel activity. Conditions appropriate for such treatment include those set forth above. The compounds identified are formulated in conventional ways as set forth in Remington's "Pharmaceutical Sciences," latest edition, Mac Publishing Co., Easton, PA. Modes of administration are those appropriate for the condition to be treated and are within the ordinary skill of the practitioner.

In addition, the regulation of expression of T-type calcium channels can be achieved by constructing expression systems encoding antisense sequences or sequences designed for triplex binding to interrupt the expression of nucleotide sequences encoding the T-type calcium channels of the invention.

DNA fragments with sequences given by SEQ ID NOS. 13-17 and 19, or polynucleotides with the complete coding sequences as given by SEQ ID NOS. 23, 25



and 27 or Figure 6 or distinctive portions thereof which do not exhibit non-discriminatory levels of homology with other types of calcium channel subunits may also be used for mapping the distribution of  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  calcium channel subunits within a tissue sample. This method follows normal histological procedures using a nucleic acid probe, and generally involves the steps of exposing the tissue to a reagent comprising a directly or indirectly detectable label coupled to a selected DNA fragment, and detecting reagent that has bound to the tissue. Suitable labels include fluorescent labels, enzyme labels, chromophores and radio-labels.

### Heterologous Expression of Mammalian T-type Calcium Channels in Cells

#### A. Transient Transfection in Mammalian Cells

Host cells, such as human embryonic kidney cells, HEK 293 (ATCC# CRL 1573) are grown in standard DMEM medium supplemented with 2 mM glutamine and 10% fetal bovine serum. HEK 293 cells are transfected by a standard calcium-phosphate-DNA co-precipitation method using a full-length mammalian  $\alpha_1$  T-type calcium channel cDNA (for example, SEQ. ID. NO. 27) in a vertebrate expression vector (for example see Current protocols in Molecular Biology). The  $\alpha_{1I}$  calcium channel cDNA may be transfected alone or in combination with other cloned subunits for mammalian calcium channels, such as  $\alpha_{2\delta}$  and  $\beta$  or  $\gamma$  subunits, and also with clones for marker proteins such as the jellyfish green fluorescent protein.

Electrophysiological Recording: After an incubation period of from 24 to 72 hrs the culture medium is removed and replaced with external recording solution (see below). Whole cell patch clamp experiments are performed using an Axopatch 200B amplifier (Axon Instruments, Burlingame, CA) linked to an IBM compatible personal computer equipped with pCLAMP software. Microelectrodes are filled with 3 M CsCl and have typical resistances from 0.5 to 2.5 Mohms. The external recording solution is 2 mM  $\text{BaCl}_2$ , 1 mM  $\text{MgCl}_2$ , 10 mM HEPES, 40 mM TEACl, 10 mM Glucose, 92 mM CsCl, (pH 7.2). The internal pipette solution is 105 mM CsCl, 25 mM TEACl, 1 mM  $\text{CaCl}_2$ , 11 mM EGTA, 10 mM HEPES (pH 7.2). Currents are typically elicited from a holding potential of -100 mV to various test potentials. Data are filtered at 1 kHz and recorded directly on the hard-drive of a personal computer. Leak subtraction is carried out on-line

using a standard P/5 protocol. Currents are analyzed using pCLAMP versions 5.5 and 6.0. Macroscopic current-voltage relations are fitted with the equation  $I = \frac{1}{1 + \exp(-(V_m - V_h)/S)} \times G - (V_m - E_{rev})$ , where  $V_m$  is the test potential,  $V_h$  is the voltage at which half of the channels are activated, and  $S$  reflects the steepness of the activation curve and is an indication of the effective gating charge movement. Inactivation curves are normalized to 1 and fitted with  $I = (1 / (1 + \exp((V_m - V_h)/S)))$  with  $V_m$  being the holding potential. Single channel recordings are performed in the cell-attached mode with the following pipette solution (in mM): 100 BaCl<sub>2</sub>, 10 HEPES, pH 7.4 and bath solution: 100 KCl, 10 EGTA, 2 MgCl<sub>2</sub>, 10 HEPES, pH 7.4.

#### B. Transient Transfection in *Xenopus* Oocytes

Stage V and VI *Xenopus* oocytes are prepared as described by Dascal, *et al* (1986), Expression and modulation of voltage-gated calcium channels after RNA injection into *Xenopus* oocytes. Science 231:1147-1150. After enzymatic dissociation with collagenase, oocytes nuclei are microinjected with the human  $\alpha_{1I}$  calcium channel cDNA expression vector construct (approximately 10 ng DNA per nucleus) using a Drummond nanoject apparatus. The  $\alpha_{1I}$  calcium channel may be injected alone, or in combination with other mammalian calcium channel subunit cDNAs, such as the  $\alpha_2\delta$  and  $\beta_{1b}$  and  $\gamma$  subunits. After incubation from 48 to 96 hrs macroscopic currents are recorded using a standard two microelectrode voltage-clamp (Axoclamp 2A, Axon Instruments, Burlingame, CA) in a bathing medium containing (in mM): 40 Ba(OH)<sub>2</sub>, 25 TEA-OH, 25 NaOH, 2 CsOH, 5 HEPES (pH titrated to 7.3 with methane-sulfonic acid). Pipettes of typical resistance ranging from 0.5 to 1.5 Mohms are filled with 2.8M CsCl, 0.2M CsOH, 10mM HEPES, 10mM BAPTA free acid. Endogenous Ca (and Ba) - activated Cl currents are suppressed by systematically injecting 10-30 nl of a solution containing 100mM BAPTA-free acid, 10mM HEPES (pH titrated to 7.2 with CsOH) using a third pipette connected to a pneumatic injector. Leak currents and capacitive transients are subtracted using a standard P/5 procedure.

Construction of Stable Cell Lines Expressing Mammalian T-type Calcium Channels

Mammalian cell lines stably expressing human  $\alpha_{11}$  calcium channels are constructed by transfecting the  $\alpha_{11}$  calcium channel cDNA into mammalian cells such as HEK 293 and selecting for antibiotic resistance encoded for by an expression vector.

5 Briefly, a full-length mammalian T-type calcium channel  $\alpha_1$  subunit cDNA (for example SEQ. ID NO. 27) subcloned into a vertebrate expression vector with a selectable marker, such as the pcDNA3 (InvitroGen, San Diego, CA), is transfected into HEK 293 cells by calcium phosphate coprecipitation or lipofection or electroporation or other method according to well known procedures (Methods in Enzymology, Volume 185, Gene  
10 Expression Technology (1990) Edited by Goeddel, D.V.). The  $\alpha_{11}$  calcium channel may be transfected alone, or in combination with other mammalian calcium channel subunit cDNAs, such as the  $\alpha_2$ - $\delta$  and  $\beta_1$ b subunits, either in a similar expression vector or other type of vector using different selectable markers. After incubation for 2 days in nonselective conditions, the medium is supplemented with Geneticin (G418) at a  
15 concentration of between 600 to 800 ug/ml. After 3 to 4 weeks in this medium, cells which are resistant to G418 are visible and can be cloned as isolated colonies using standard cloning rings. After growing up each isolated colony to confluency to establish cell lines, the expression of  $\alpha_{11}$  calcium channels can be determined at with standard gene expression methods such as Northern blotting, RNase protection and reverse-transcriptase  
20 PCR.

The functional detection of  $\alpha_{11}$  calcium channels in stably transfected cells can be examined electrophysiologically, such as by whole patch clamp or single channel analysis (see above). Other means of detecting functional calcium channels include the use of radiolabeled  $^{45}\text{Ca}$  uptake, fluorescence spectroscopy using calcium sensitive dyes  
25 such as FURA-2, and the binding or displacement of radiolabeled ligands that interact with the calcium channel.

Example 1

Partial Rat and Human Subunits

In order to recover mammalian sequences for novel calcium channels, the 567 base pair partial length human brain  $\alpha_{1I}$  cDNA described in WO 98/3801 was gel-purified, radio-labeled with  $^{32}\text{P}$  dATP and dCTP by random priming (Feinberg, *et al.*, 1983, *Anal. Biochem.* 132: 6-13) and used to screen a rat brain cDNA library constructed in the phase vector Lambda Zapp II. (Snutch *et al.*, 1990, *Proc Natl Acad Sci (USA)* 87: 3391-3395). Screening was carried out at 62°C in 5XSSPE (1XSSPE= 0.18 M NaCl; 1mM EDTA; 10 mM sodium phosphate, pH=7.4 0.3% SDS, 0.2 mg/ml denatured salmon sperm DNA). Filters were washed at 62°C in 0.2X SSPE/0.1% SDS. After three rounds of screening and plaque purification, positive phages were transformed into Bluescript phagemids (Stratagene, La Jolla, CA) by *in vivo* excision.

Double stranded DNA sequencing on the recombinant phagemids was performed using a modified dideoxynucleotide protocol (Biggin *et al.*, 1983, *Proc Natl Acad Sci (USA)* 80:3963-3965) and Sequenase version 2.1 (United States Biochemical Corp.). DNA sequencing identified three distinct classes of calcium channel  $\alpha_1$  subunits: designated as  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  calcium channel subunits.

For each class of calcium channel  $\alpha_1$  subunit, the largest cDNA was completely sequenced and determined to represent only a portion of the predicted calcium channel coding region. In order to isolate the remaining portions of  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channel subunits, the  $\alpha_{1G}$  clone was digested with HindIII and SpeI. The resulting 540 base pair fragment was gel purified, radiolabeled with  $^{32}\text{P}$  dATP and dCTP by random priming and used to rescreen the rat brain cDNA library as described above. The sequence of the 540 base pair  $\alpha_{1G}$  screening probe used is given by SEQ. ID NO. 29. Other sequences spanning regions of distinctiveness within the sequences for the subunits could also be employed.

Double-stranded DNA sequencing of the purified recombinant phagemids showed that additional  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  calcium channel subunit cDNAs overlapped with the original partial length cDNAs and together encoded complete protein coding regions as well as portions of their respective 5' and 3' non-coding untranslated regions.

To recover further human sequences for the novel  $\alpha_{1G}$  and  $\alpha_{1H}$  calcium channels, the 567 base pair partial length human brain  $\alpha_{1I}$  cDNA (SEQ. ID. NO: 19) was radio-labeled with  $^{32}P$  dATP and dCTP by random priming and used to screen a commercial human thalamus cDNA library (Clontech). Hybridization was performed overnight at 65°C in 6 X SSPE; 0.3% SDS; 5X Denhardt's. Filters were washed at 65°C in 0.1 X SSPE/ 0.3% SDS. After four rounds of screening and plaque purification, positive phages were selected, DNA prepared and the insert cDNA excised from the lambda vector by digestion with Eco R1 restriction enzyme. The excised cDNA was subcloned into the plasmid Bluescript KS (Stratagene, La Jolla, CA) and the DNA sequence determined using a modified dideoxynucleotide protocol and Sequence version 2.1. The partial length  $\alpha_{1G}$  cDNA isolated consisted of 2212 base pairs of which 279 base pairs were 5' noncoding and 1,933 base pairs were coding region representing 644 amino acids (SEQ. ID NOS. 30, 31). The partial  $\alpha_{1H}$  cDNA isolated consisted of 1,608 base pairs of which 53 base pairs were 5' noncoding and 1,555 were coding region representing 518 amino acids (SEQ. ID NOS. 32, 33).

To recover further human sequences for the novel  $\alpha_{1I}$  calcium channel, the full-length rat brain  $\alpha_{1I}$  cDNA (SEQ. ID. NO: 27) (See Example 2) was radio-labeled  $^{32}P$  dATP and dCTP by random priming and used to screen a commercial human hippocampus cDNA library (Stratagene). Hybridization was performed overnight at 65 °C in 6 X SSPE; 0.3% SDS; 5X Denhardt's. Filters were washed at 65 °C in 0.1 X SSPE/ 0.3% SDS. After four rounds of screening and plaque purification, positive phages were transformed into Bluescript phagemids (Stratagene, LA Jolla, CA) by *in vitro* excision. The excised cDNA DNA sequence was determined using a modified dideoxynucleotide protocol and Sequenase version 2.1. The partial  $\alpha_{1I}$  cDNA isolated consisted of 1,080 base pairs of coding region representing 360 amino acids (SEQ. ID NOS. 34, 35).

### Example 2

#### Full Length Rat Subunits

Double-stranded DNA sequencing of the purified recombinant phagemids from rat brain showed that additional  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channel cDNAs overlapped with the original partial length cDNAs and together encoded complete protein coding regions as

well as portions of their respective 5' and 3' non-coding untranslated regions. (SEQ. ID NOS. 23 and 27, respectively) In addition to the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channel classes, DNA sequencing of the recombinant phagemids also identified a third class of calcium channel  $\alpha_1$  subunit: designated as the  $\alpha_{1H}$  calcium channel subunit. The partial length  $\alpha_{1H}$  calcium channel cDNAs overlapped and together encoded a complete  $\alpha_{1H}$  coding region as well as portions of the 5' and 3' untranslated regions (SEQ. ID. NO. 25).

Electrophysiological studies were performed on transiently-transfected human embryonic kidney cells (HEK-tsa201) prepared using the general protocol above. Transfection was carried out by standard calcium phosphate precipitation. (Okayama *et al.*, 1991, *Methods in Molec. Biol.*, Vol. 7, ed. Murray, E.J.). For maintenance, HEK-tsa201 cells were cultured until approximately 70% confluent, the media removed and cells dispersed with trypsin and gentle trituration. Cells were then diluted 1:10 in culture medium (10% FBS, DMEM plus L-glutamine, pen-strep) warmed to 37°C and plated onto tissue culture dishes. For transient transfection, 0.5 mM  $\text{CaCl}_2$  was mixed with a total of 20  $\mu\text{g}$  of DNA (consisting of 3 $\mu\text{g}$  of either rat brain  $\alpha_{1G}$  or  $\alpha_{1I}$  calcium channel cDNA, 2  $\mu\text{g}$  of CD8 plasmid marker, and 15  $\mu\text{g}$  of Bluescript plasmid carrier DNA). The DNA mixture was mixed thoroughly and then slowly added dropwise to 0.5 ml of 2 times HeBS (274 mM NaCl, 20mM D-glucose, 10mM KCl, 1.4 mM  $\text{Na}_2\text{HPO}_4$ , 40 mM Hepes (pH=7.05). After incubation at room temperature for 20 min, 100  $\mu\text{l}$  of the DNA mixture was slowly added to each dish of HEK-tsa201 cells and then incubated at 37°C for 24 to 48 hours in a tissue culture incubator (5%  $\text{CO}_2$ ).

Positive transfectant cells were identified visually by addition of 1  $\mu\text{l}$  of mouse CD8 (Lyt2) Dynabeads directly to the recording solution and gentle swirling to mix. Whole cell patch clamp readings were carried out with an Axopatch 200A amplifier (Axon Instruments) as described previously. (Zamponi *et al.*, 1997, *Nature* 385: 442-446). The external recording solution was 2 mM  $\text{CaCl}_2$ , 1 mM  $\text{MgCl}_2$ , 10 mM HEPES, 40 mM TEA-Cl, 10 mM glucose, 92 mM CsCl, pH=7.2 with TEA-hydroxide. The internal pipette solutions was 105 mM CsCl, 25 mM TEA-Cl, 1mM  $\text{CaCl}_2$ , 11 mM EGTA, 10 mM HEPES, pH 7.2 with NaOH.

For determination of current-voltage (I-V) relationships, cells were held at a resting potential of -100 mV and then stepped to various depolarizing test potentials. For

steady-state inactivation, cells were held at various potentials for 20 sec. and currents recorded during a subsequent test pulse to the peak potential of the I-V. Leak currents and capacitive transients were subtracted using a P/5 procedure.

5 Figs. 1-4 show the results obtained for HEK cells transfected with and expressing the cDNA of sequences ID Nos. 23 and 27, which correspond to the subunits designated as  $\alpha_{1G}$  and  $\alpha_{1I}$ . Figs. 1A and B and 2A and B shows a comparison of the waveforms and current- voltage relationship for the two channel subunit types. In the presence of recording solution containing 2mM  $\text{Ca}^{2+}$ , both the  $\alpha_{1G}$  and  $\alpha_{1I}$  channel subunits exhibit activation properties consistent with native T-type calcium currents. Figs. 1A and 2A 10 show the subunit calcium current from a cell held at -120 mV and depolarized to a series of test potentials. Figs. 1B and 2B show the magnitude of the calcium current. From a holding potential of -110 mV, both channel first activate at approximately -70 mV and peak currents are obtained between -40 and -50 mV. Upon depolarization to various test potentials, the current waveforms of the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channels exhibit an 15 overlapping pattern characteristic of native T-type channels in nerve, muscle and endocrine cells.

Fig. 3 shows steady-state inactivation profiles for the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channels in HEK 293 cells transiently transformed with full length cDNAs (SEQ ID NOS. 23 or 27) for  $\alpha_{1G}$  or  $\alpha_{1I}$  subunits. Steady state inactivation properties were 20 determined by stepping from -120 mV to prepulse holding potentials between -120 mV and -50 mV for 15 sec.. prior to a test potential of -30 mV. The data are plotted as normalized whole cell current versus prepulse holding potential and show that  $\alpha_{1G}$  exhibits a  $V_{50}$  of approximately -85 mV and  $\alpha_{1I}$  a  $V_{50}$  of approximately -93 mV. Thus, consistent with native T-type calcium channels, both of the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channels 25 exhibit pronounced steady-state inactivation at negative potentials.

Figs. 4A-C show a comparison of the voltage-dependent deactivation profiles of the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channels. HEK 293 cells were transiently transfected with either an  $\alpha_{1G}$  or  $\alpha_{1I}$  subunit cDNA (SEQ. ID NO. 23 or 27). The deactivation properties of  $\alpha_{1G}$  were determined by stepping from a holding potential of -100 mV to -40mV for 9 msec, 30 and then to potentials between -120 mV and -45 mV. The deactivation properties of  $\alpha_{1I}$  were determined by stepping from a holding potential of -100 mV to -40 mV for 20

msec, and then to potentials between -120 mV and -45 mV. Both channels exhibit slow deactivation kinetics compared to typical high-threshold channels, and is consistent with the  $\alpha_{1G}$  and  $\alpha_{1I}$  subunits being subunits for T-type calcium channels

### Example 3

#### 5      Cloning of a Full Length cDNA for the Human $\alpha_{1G}$ T-Type Calcium Channel Subunit

##### Materials and Methods:

A full length cDNA encoding the human  $\alpha_{1G}$  subunit was constructed from 5 overlapping clones (Figure 1B) isolated from a human thalamus cDNA library constructed in  $\lambda$ gt11 vector (Clontech, Cat#HL5009b).s

10          Three  $\lambda$ gt11 cDNA clones were isolated by conventional filter hybridization.

Clone 1 was identified by hybridization to a 567 bp cDNA probe (SEQ. ID. NO: 19) containing the transmembrane region S4 to S6 of domain I of the previously cloned human neuronal  $\alpha_{1I}$  T-type calcium channel subunit. Clones HG10-1112 and HG5-1211 were identified by hybridization to a 1265 bp cDNA probe of the rat  $\alpha_{1H}$  T-type calcium channel subunit spanning domain II and part of the II-III intracellular loop. cDNA probes were  $^{32}$ P-dCTP labeled by random priming using a Multiprime DNA labeling system (Amersham Pharmacia). Plaque lifts using H-bond nylon membranes were done in duplicate following the standard protocols supplied by manufacturer (Amersham Pharmacia). Hybridization was performed for at least 16 hrs at 65°C for clone 1 and for at least 16 hrs at 58°C, clones HG10-1112 and HG5-1211. Membranes were washed in 0.1X SSC/0.3% SDS at 62°C for clone 1 and 0.2X SSC/0.1% SDS at 58°C clones HG10-1112 and HG5-1211. Blots were exposed to BioMax MS Kodak film with Kodak HE intensifying screens for at least 48 hrs at -80°C. Double positive plaques were isolated and re-screened to isolate single clones according to the procedure above.

25          Bacteriophage DNA's were then isolated according to the  $\lambda$ gt11 library User Manual (Clontech). Clone 1 cDNA insert was excised with EcoRI (NEB) and subcloned into pBluescriptKS (Stratagene). Clones HG10-1112 and HG5-1211 cDNA inserts were excised from  $\lambda$ DNA with Not I (NEB) and subcloned into the Not I site of pBluescriptKS. Plasmids with cDNA inserts were transformed by electroporation into



XL-I *E. coli* host strain bacteria and sequenced using universal reverse and forward primers according to Sanger double stranded DNA sequencing method in combination with automatic sequencing ABI 100 PRISM model 377 Version 3.3 (PE Biosystems).

Clone 1 was identified as a human  $\alpha_{1G}$  subunit containing the 5'UTR and 1933 bp of the in-frame coding region, including part of the intracellular I-II loop. Clone HG10-1112 was identified as a human  $\alpha_{1G}$  subunit of 3915 bp, spanning DomainI (S5-S6) to the III-IV loop. Clone HG5-1211 was identified as human  $\alpha_{1G}$  subunit of 3984 bp containing the I-II linker and C-terminus.

For expression in HEK cells, removal of 5' UTR from clone 1 was achieved by replacing 5'UTR DNA fragment flanked by Hind III/SacII restriction sites with 5'end - 291 bp cDNA fragment, containing translation start site and an incorporated Hind III site for subsequent cloning into pcDNA3.1 (Invitrogen). Following PCR conditions were used: 94°C -30 sec, 45°C -30 sec, 72°C -30 sec for 5 cycles and followed by 94°C -30 sec, 48°C -30 sec, 72°C -30 sec for 20 cycles (Bio-rad Gene Cyclor). The cDNA fragment was subcloned into p-Gem-T-Easy plasmid vector (Promega) and the DNA sequence determined.

The remaining region of the 3'  $\alpha_{1G}$  subunit cDNA was obtained using the PCR method on a human thalamus cDNA library with primers MD19-sense (5'GCG TGG AGC TCT TTG GAG 3') and G26- antisense (5' GCA CCC AGT GGA GAA AGG TG 3'). The PCR protocol used was 94°C -30 sec, 58°C -30 sec, 72°C -30 sec for 25 cycles (Bio-rad Gene Cyclor). A cDNA fragment of 1617 bp was subcloned into p-Gem-T-Easy plasmid vector (Promega) and sequenced. The 3'PCR cDNA was identified as a human  $\alpha_{1G}$  subunit spanning from Domain IV-S5 to the carboxyl terminus including the stop codon.

Unique restriction sites (Figures 5A and B) of the partial cDNA clones were used to construct the full length human  $\alpha_{1G}$  T-type calcium channel in pcDNA3.1 Zeo (+) (Invitrogen) mammalian expression vector.

The complete nucleotide and amino acid sequences are shown in Figure 6.

In order to determine the functional properties of the human  $\alpha_{1G}$  channel standard calcium-phosphate transfection was used to transiently express the channel in HEK ts201

cells. Cells were cultured in Dulbecco's modified eagle medium (DMEM) supplemented with 10% fetal bovine serum, 200 U/ml penicillin and 0.2 mg/ml streptomycin at 37°C with 5% CO<sub>2</sub>. At 85% confluency cells were split with 0.25% trypsin/1 mM EDTA and plated at 10% confluency on glass coverslips. At 12 hours the medium was replaced and the cells transiently transfected using a standard calcium phosphate protocol and the  $\alpha_{1G}$  calcium channel cDNA. Fresh DMEM was supplied and the cells transferred to 28°C/5% CO<sub>2</sub>. Cells were incubated for 1 to 2 days prior to whole cell recording. Whole cell patch recordings were performed using an Axopatch 200B amplifier (Axon Instruments) linked to an IBM compatible personal computer equipped with pCLAMP version 7.0 software. The intrapipette solution contained (in mM): 105 CsCl, 25 CsCl, 1 CaCl<sub>2</sub>, 11 EGTA, 10 HEPES, pH 7.2. The extracellular solution contained (in mM): 40 TEA-Cl, 2 CaCl<sub>2</sub>, 1 MgCl<sub>2</sub>, 92 CsCl, 10 glucose, 10 HEPES, pH 7.2.

Figure 7 shows that the human  $\alpha_{1G}$  cDNA encodes a calcium channel with typical properties of a T-type current. The left panel illustrates representative current traces obtained from a holding potential of -100 mV to test pulses potentials of -90 mV to +20 mV. The traces show a typical crossover pattern and considerable inactivation during the test pulse, both of which are consistent with native T-type channels. The right panel shows a plot of the peak whole current at various test potentials and indicates that the human  $\alpha_{1G}$  cDNA first activates near -60 mV with maximal current near -40 mV, which is also consistent with native low-threshold T-type calcium channels.

Claims

1. A DNA molecule which comprises an expression cassette wherein said expression cassette comprises a nucleotide sequence encoding a T-type calcium channel  $\alpha_1$  subunit, said encoding sequence operably linked to control sequences to effect its expression.

2. The DNA molecule of claim 1 wherein said  $\alpha_1$  subunit is  $\alpha_{1G}$ ,  $\alpha_{1H}$ , or  $\alpha_{1I}$ .

3. The DNA molecule of claim 2 wherein said  $\alpha_1$  subunit is derived from a mammal.

4. Recombinant host cells modified to contain the DNA molecule of any of claims 1-3.

5. The cells of claim 4 which are mammalian cells.

6. A method to effect production of a functional calcium channel which method comprises culturing the cells of claim 4 or 5 under conditions wherein said functional calcium channels are produced.

7. A method to identify a compound which is a modulator for T-type mammalian calcium channels, which method comprises contacting the cells employed in the method of claim 6 with said compound and assessing the effect of said compound on said cells.

8. A T-type calcium channel modulator identified by the method of claim 7.

9. A method to treat conditions characterized by undesirable levels of T-type calcium channel activity which method comprises administering to a subject in need of such treatment an effective amount of the modulator of claim 8.

10. The method of claim 9 wherein said condition is cardiac hypertrophy, cardiac arrhythmia, hypertension, a sleep disorder, or epilepsy.

11. A DNA molecule which comprises an expression system for a nucleotide sequence which is complementary to the nucleotide sequence encoding a T-type calcium channel  $\alpha_1$  subunit or which forms a triple helix with DNA comprising said encoding sequence.

12. A method to treat a condition characterized by an undesirable level of T-type calcium channel activity which method comprises administering to a subject in need of such treatment an effective amount of the DNA molecule of claim 11.

13. The method of claim 12 wherein said condition is cardiac hypertrophy, cardiac arrhythmia, hypertension, a sleep disorder, or epilepsy.

14. An oligonucleotide which consists essentially of a nucleotide sequence characteristic of a T-type calcium channel  $\alpha_1$  subunit, said oligonucleotide coupled to or comprising a detectable label.

15. A method to map the distribution of T-type calcium channels in a tissue which method comprises contacting said tissue with the oligonucleotide of claim 14.

16. Antibodies specifically immunoreactive with the extracellular portions of a T-type calcium channel.

17. A method to map the distribution of T-type calcium channels in a tissue which method comprises contacting said tissue with the antibodies of claim 16.

ABSTRACT OF THE DISCLOSURE

Sequences and partial sequences for three types of mammalian (human and rat sequences identified) T-type calcium channel subunits which we have labeled as the  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits are provided. Knowledge of the sequence of these calcium channel permits the localization and recovery of the complete sequence from human cells, and the development of cell lines which express the novel calcium channels of the invention. These cells may be used for identifying compounds capable of acting as agonists or antagonists to the calcium channels.

5

$\alpha_{1G}$

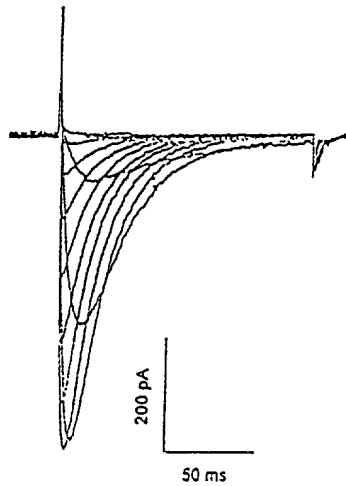


Fig. 1A

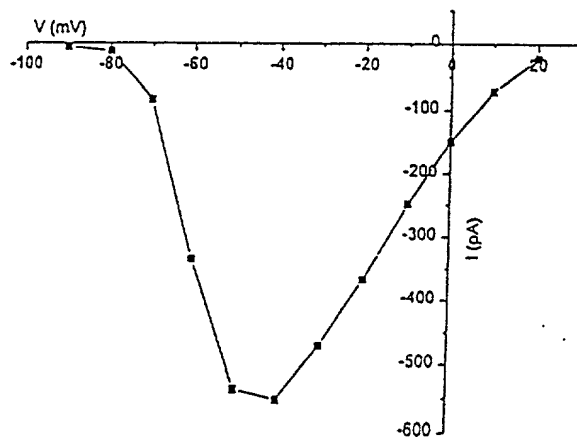


Fig. 1B

$\alpha_{11}$

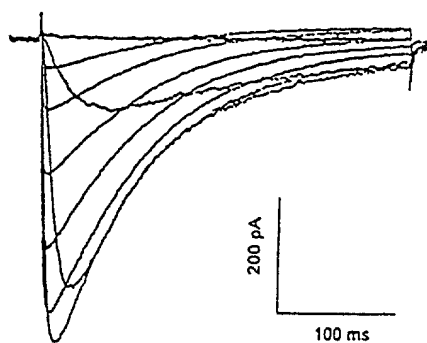


Fig. 2A

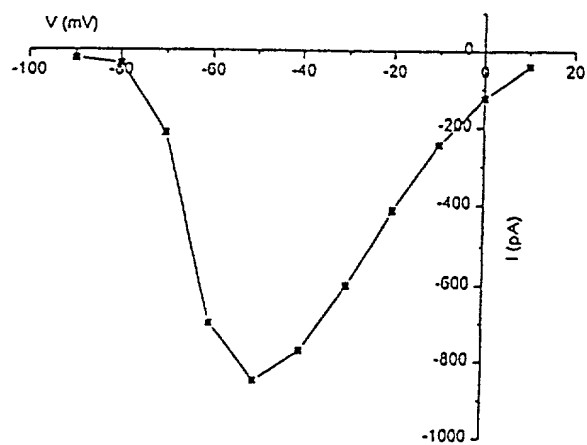


Fig. 2 B

# Steady-state inactivation

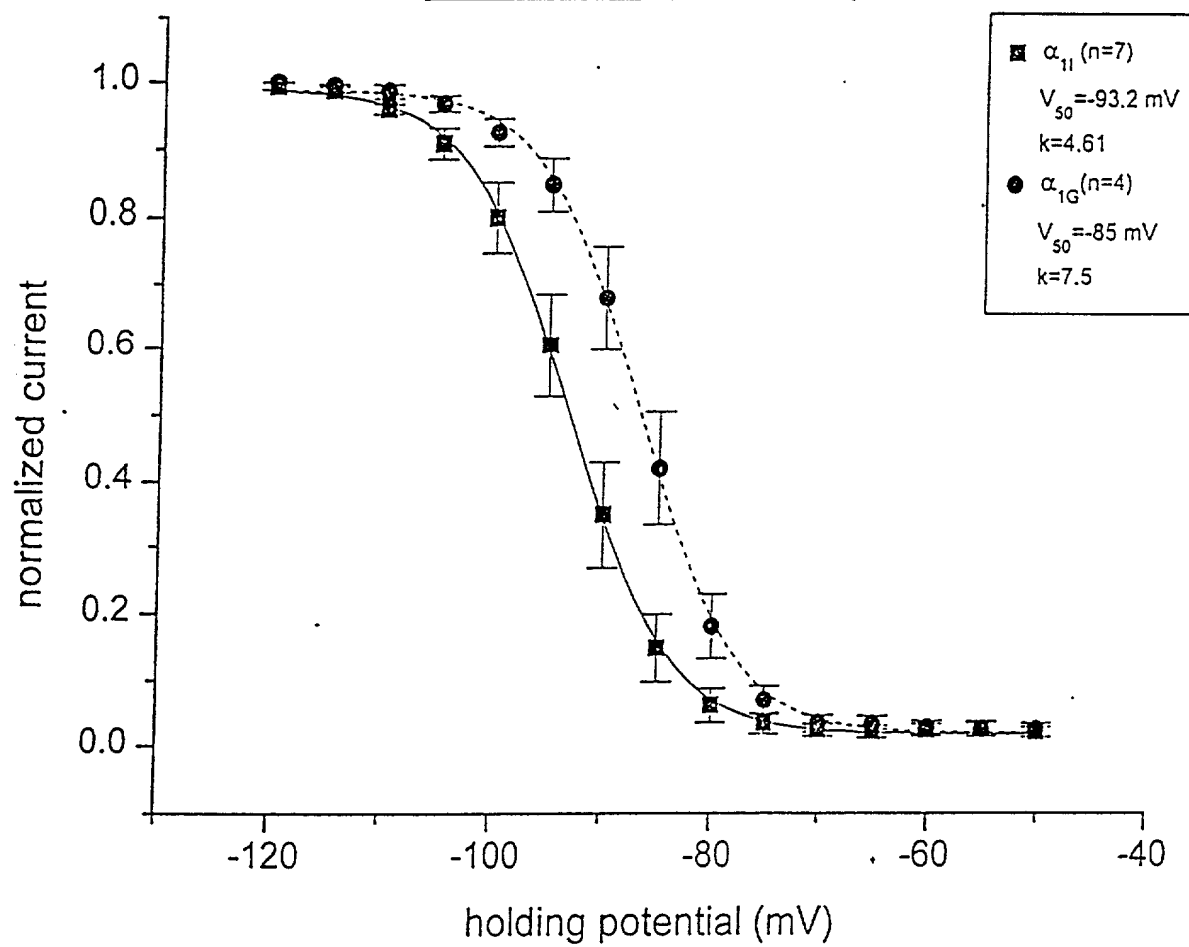


Fig. 3



Deactivation

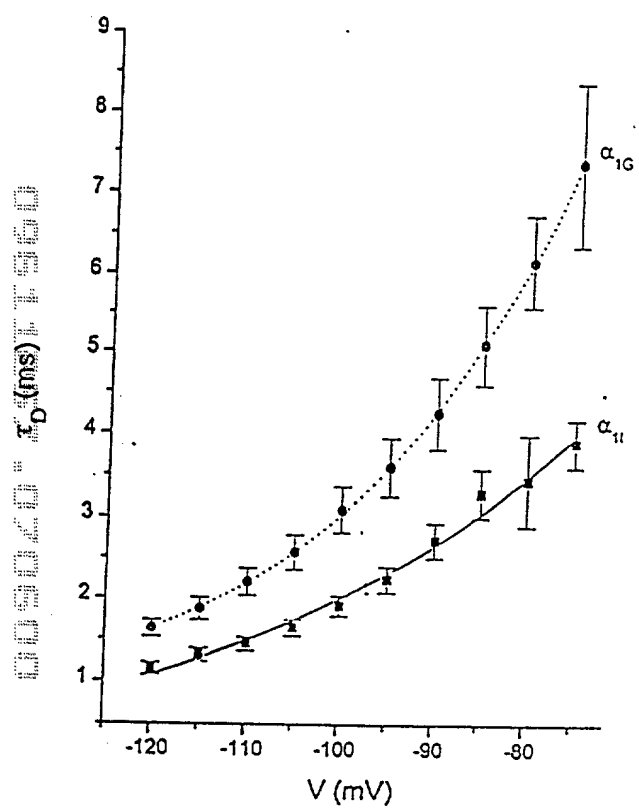
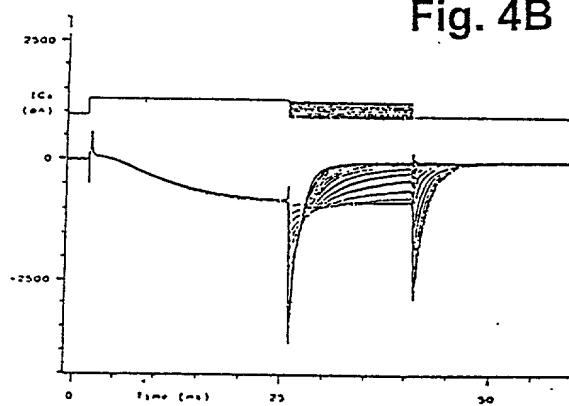


Fig. 4A

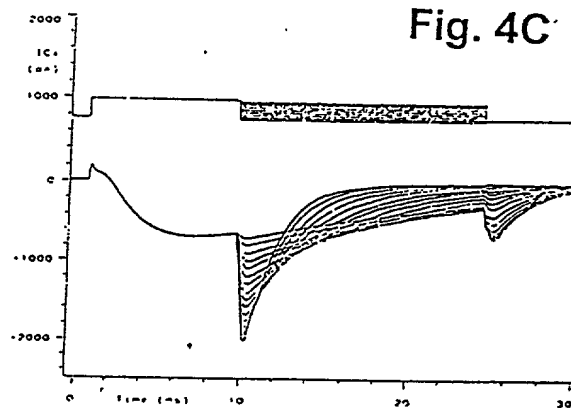
$\alpha_{1I}$

Fig. 4B



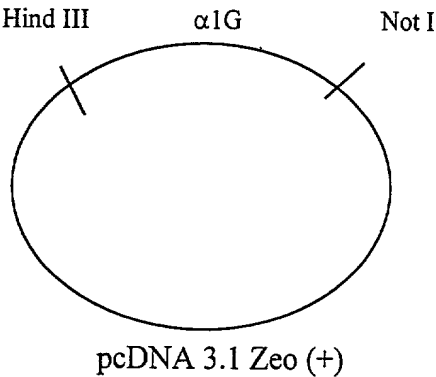
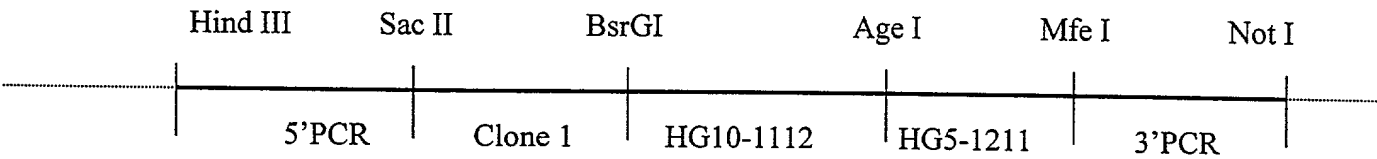
$\alpha_{1G}$

Fig. 4C



5

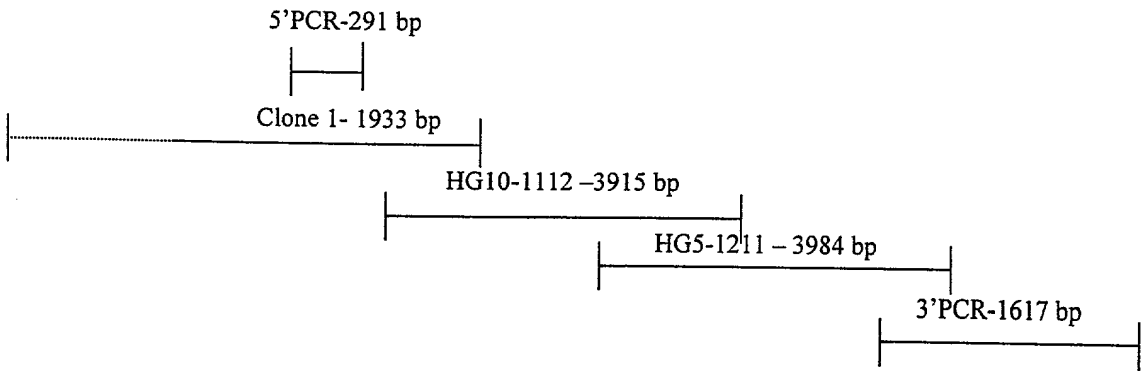
**Figure 1A.  $\alpha$ 1G cDNA construct**



pcDNA 3.1 Zeo (+)

5

**Figure 1B.  $\alpha$ 1G cDNA CLONES**



### Human $\alpha 1$ G T-type calcium channel cDNA

1 aagcttgcttgcccctctccggatcgcccggggccccggctggccagagg ATG GAC GAG GAG GAG GAT GGA 71  
1 M D E E E D G 7  
72 GCG GGC GCC GAG GAG TCG GGA CAG CCC CGG AGC TTC ATG CGG CTC AAC GAC CTG TCG GGG 131  
8 A G A E E S G Q P R S F M R L N D L S G 27  
132 GCC GGG GGC CGG CCG GGG CCG GGG TCA GCA GAA AAG GAC CCG GGC AGC GCG GAC TCC GAG 191  
28 A G G R P G G S A E K D P G S A D S E 47  
192 GCG GAG GGG CTG CCG TAC CCG GCG CTG GCC CCG GTG GTT TTC TTC TAC TTG AGC CAG GAC 251  
48 A E G L P Y P A L A P V V F F Y L S Q D 67  
252 AGC CGC CCG CGG AGC TGG TGT CTC CGC ACG GTC TGT AAC CCC TGG TTT GAG CGC ATC AGC 311  
68 S R P R S W C L R T V C N P W F E R I S 87  
312 ATG TTG GTC ATC CTT CTC AAC TGC GTG ACC CTG GGC ATG TTC CGG CCA TGC GAG GAC ATC 371  
88 M L V I L L N C V T L G M F R P C E D I 107  
372 GCC TGT GAC TCC CAG CGC TGC CGG ATC CTG CAG GCC TTT GAT GAC TTC ATC TTT GCC TTC 431  
108 A C D S Q R C R I L Q A F D D F I F A F 127  
432 TTT GCC GTG GAG ATG GTG GTG AAG ATG GTG GCC TTG GGC ATC TTT GGG AAA AAG TGT TAC 491  
128 F A V E M V V K M V A L G I F G K K C Y 147  
492 CTG GGA GAC ACT TGG AAC CGG CTT GAC TTT TTC ATC GTC ATC GCA GGG ATG CTG GAG TAC 551  
148 L G D T W N R L D F F I V I A G M L E Y 167  
552 TCG CTG GAC CTG CAG AAC GTC AGC TTC TCA GCT GTC AGG ACA GTC CGT GTG CTG CGA CCG 611  
168 S L D L Q N V S F S A V R T V R V L R P 187  
612 CTC AGG GCC ATT AAC CGG GTG CCC AGC ATG CGC ATC CTT GTC ACG TTG CTG CTG GAT ACG 671  
188 L R A I N R V P S M R I L V T L L L D T 207  
672 CTG CCC ATG CTG GGC AAC GTC CTG CTG CTC TGC TTC TTC GTC TTC TTC ATC TTC GGC ATC 731  
208 L P M L G N V L L L C F F V F F I F G I 227  
732 GTC GGC GTC CAG CTG TGG GCA GGG CTG CTT CGG AAC CGA TGC TTC CTA CCT GAG AAT TTC 791  
228 V G V Q L W A G L L R N R C F L P E N F 247  
792 AGC CTC CCC CTG AGC GTG GAC CTG GAG CGC TAT TAC CAG ACA GAG AAC GAG GAT GAG AGC 851  
248 S L P L S V D L E R Y Y Q T E N E D E S 267  
852 CCC TTC ATC TGC TCC CAG CCA CGC GAG AAC GGC ATG CGG TCC TGC AGA AGC GTG CCC ACG 911  
268 P F I C S Q P R E N G M R S C R S V P T 287  
912 CTG CGC GGG GAC GGG GGC GGT GGC CCA CCT TGC GGT CTG GAC TAT GAG GCC TAC AAC AGC 971  
288 L R G D G G G P P C G L D Y E A Y N S 307  
972 TCC AGC AAC ACC ACC TGT GTC AAC TGG AAC CAG TAC TAC ACC AAC TGC TCA GCG GGG GAG 1031  
308 S S N T T C V N W N Q Y Y T N C S A G E 327  
1032 CAC AAC CCC TTC AAG GGC GCC ATC AAC TTT GAC AAC ATT GGC TAT GCC TGG ATC GCC ATC 1091  
328 H N P F K G A I N F D N I G Y A W I A I 347  
1092 TTC CAG GTC ATC ACG CTG GAG GGC TGG GTC GAC ATC ATG TAC TTT GTG ATG GAT GCT CAT 1151  
348 F Q V I T L E G W V D I M Y F V M D A H 367  
1152 TCC TTC TAC AAT TTC ATC TAC TTC ATC CTC CTC ATC ATC GTG GGC TCC TTC TTC ATG ATC 1211  
368 S F Y N F I Y F I L L I I V G S F F M I 387  
1212 AAC CTG TGC CTG GTG GTG ATT GCC ACG CAG TTC TCA GAG ACC AAG CAG CGG GAA AGC CAG 1271  
388 N L C L V V I A T Q F S E T K Q R E S Q 407  
1272 CTG ATG CGG GAG CAG CGT GTG CGG TTC CTG TCC AAC GCC AGC ACC CTG GCT AGC TTC TCT 1331  
408 L M R E Q R V R F L S N A S T L A S F S 427  
1332 GAG CCC GGC AGC TGC TAT GAG GAG CTG CTC AAG TAC CTG GTG TAC ATC CTT CGT AAG GCA 1391  
428 E P G S C Y E E L L K Y L V Y I L R K A 447

Figure 6

1392	GCC	CGC	AGG	CTG	GCT	CAG	GTC	TCT	CGG	GCA	GCA	GGT	GTG	CGG	GTT	GGG	CTG	CTC	AGC	AGC	1451
448	A	R	R	L	A	Q	V	S	R	A	A	G	V	R	V	G	L	L	S	S	467
1452	CCA	GCA	CCC	CTC	GGG	GGC	CAG	GAG	ACC	CAG	CCC	AGC	AGC	AGC	TGC	TCT	CGC	TCC	CAC	CGC	1511
468	P	A	P	L	G	G	Q	E	T	Q	P	S	S	S	C	S	R	S	H	R	487
1512	CGC	CTA	TCC	GTC	CAC	CAC	CTG	GTG	CAC	CAC	CAC	CAC	CAC	CAT	CAC	CAC	CAC	TAC	CAC	CTG	1571
488	R	L	S	V	H	H	L	V	H	H	H	H	H	H	H	H	H	Y	H	L	507
1572	GGC	AAT	GGG	ACG	CTC	AGG	GCC	CCC	CGG	GCC	AGC	CCG	GAG	ATC	CAG	GAC	AGG	GAT	GCC	AAT	1631
508	G	N	G	T	L	R	A	P	R	A	S	P	E	I	Q	D	R	D	A	N	527
1632	GGG	TCC	CGC	AGG	CTC	ATG	CTG	CCA	CCA	CCC	TCG	ACG	CCT	GCC	CTC	TCC	GGG	GCC	CCC	CCT	1691
528	G	S	R	R	L	M	L	P	P	P	S	T	P	A	L	S	G	A	P	P	547
1692	GGT	GGC	GCA	GAG	TCT	GTG	CAC	AGC	TTC	TAC	CAT	GCC	GAC	TGC	CAC	TTA	GAG	CCA	GTC	CGC	1751
548	G	G	A	E	S	V	H	S	F	Y	H	A	D	C	H	L	E	P	V	R	567
1752	TGC	CAG	GCG	CCC	CCT	CCC	AGG	TCC	CCA	TCT	GAG	GCA	TCC	GGC	AGG	ACT	GTG	GGC	AGC	GGG	1811
568	C	Q	A	P	P	P	R	S	P	S	E	A	S	G	R	T	V	G	S	G	587
1812	AAG	GTG	TAT	CCC	ACC	GTG	CAC	ACC	AGC	CCT	CCA	CCG	GAG	ACG	CTG	AAG	GAG	AAG	GCA	CTA	1871
588	K	V	Y	P	T	V	H	T	S	P	P	P	E	T	L	K	E	K	A	L	607
1872	GTA	GAG	GTG	GCT	GCC	AGC	TCT	GGG	CCC	CCA	ACC	CTC	ACC	AGC	CTC	AAC	ATC	CCA	CCC	GGG	1931
608	V	E	V	A	A	S	S	G	P	P	T	L	T	S	L	N	I	P	P	G	627
1932	CCC	TAC	AGC	TCC	ATG	CAC	AAG	CTG	CTG	GAG	ACA	CAG	AGT	ACA	GGT	GCC	TGC	CAA	AGC	TCT	1991
628	P	Y	S	S	M	H	K	L	L	E	T	Q	S	T	G	A	C	Q	S	S	647
1992	TGC	AAG	ATC	TCC	AGC	CCT	TGC	TTG	AAA	GCA	GAC	AGT	GGA	GCC	TGT	GGT	CCA	GAC	AGC	TGC	2051
648	C	K	I	S	S	P	C	L	K	A	D	S	G	A	C	G	P	D	S	C	667
2052	CCC	TAC	TGT	GCC	CGG	GCC	GGG	GCA	GGG	GAG	GTG	GAG	CTC	GCC	GAC	CGT	GAA	ATG	CCT	GAC	2111
668	P	Y	C	A	R	A	G	A	G	E	V	E	L	A	D	R	E	M	P	D	687
2112	TCA	GAC	AGC	GAG	GCA	GTT	TAT	GAG	TTC	ACA	CAG	GAT	GCC	CAG	CAC	AGC	GAC	CTC	CGG	GAC	2171
688	S	D	S	E	A	V	Y	E	F	T	Q	D	A	Q	H	S	D	L	R	D	707
2172	CCC	CAC	AGC	CGG	CGG	CAA	CGG	AGC	CTG	GGC	CCA	GAT	GCA	GAG	CCC	AGC	TCT	GTG	CTG	GCC	2231
708	P	H	S	R	R	Q	R	S	L	G	P	D	A	E	P	S	S	V	L	A	727
2232	TTC	TGG	AGG	CTA	ATC	TGT	GAC	ACC	TTC	CGA	AAG	ATT	GTG	GAC	AGC	AAG	TAC	TTT	GGC	CGG	2291
728	F	W	R	L	I	C	D	T	F	R	K	I	V	D	S	K	Y	F	G	R	747
2292	GGA	ATC	ATG	ATC	GCC	ATC	CTG	GTC	AAC	ACA	CTC	AGC	ATG	GGC	ATC	GAA	TAC	CAC	GAG	CAG	2351
748	G	I	M	I	A	I	L	V	N	T	L	S	M	G	I	E	Y	H	E	Q	767
2352	CCC	GAG	GAG	CTT	ACC	AAC	GCC	CTA	GAA	ATC	AGC	AAC	ATC	GTC	TTC	ACC	AGC	CTC	TTT	GCC	2411
768	P	E	E	L	T	N	A	L	E	I	S	N	I	V	F	T	S	L	F	A	787
2412	CTG	GAG	ATG	CTG	CTG	AAG	CTG	CTT	GTG	TAT	GGT	CCC	TTT	GGC	TAC	ATC	AAG	AAT	CCC	TAC	2471
788	L	E	M	L	L	K	L	L	V	Y	G	P	F	G	Y	I	K	N	P	Y	807
2472	AAC	ATC	TTC	GAT	GGT	GTC	ATT	GTG	GTC	ATC	AGC	GTG	TGG	GAG	ATC	GTG	GGC	CAG	CAG	GGG	2531
808	N	I	F	D	G	V	I	V	V	I	S	V	W	E	I	V	G	Q	Q	G	827
2532	GGC	GGC	CTG	TCG	GTG	CTG	CGG	ACC	TTC	CGC	CTG	ATG	CGT	GTG	CTG	AAG	CTG	GTG	CGC	TTC	2591
828	G	G	L	S	V	L	R	T	F	R	L	M	R	V	L	K	L	V	R	F	847
2592	CTG	CCG	GCG	CTG	CAG	CGG	CAG	CTG	GTG	GTG	CTC	ATG	AAG	ACC	ATG	GAC	AAC	GTG	GCC	ACC	2651
848	L	P	A	L	Q	R	Q	L	V	V	L	M	K	T	M	D	N	V	A	T	867
2652	TTC	TGC	ATG	CTG	CTT	ATG	CTC	TTC	ATC	TTC	ATC	TTC	AGC	ATC	CTG	GGC	ATG	CAT	CTC	TTC	2711
868	F	C	M	L	L	M	L	F	I	F	I	F	S	I	L	G	M	H	L	F	887
2712	GGC	TGC	AAG	TTT	GCC	TCT	GAG	CGG	GAT	GGG	GAC	ACC	CTG	CCA	GAC	CGG	AAG	AAT	TTT	GAC	2771
888	G	C	K	F	A	S	E	R	D	G	D	T	L	P	D	R	K	N	F	D	907
2772	TCC	TTG	CTC	TGG	GCC	ATC	GTC	ACT	GTC	TTT	CAG	ATC	CTG	ACC	CAG	GAG	GAC	TGG	AAC	AAA	2831
908	S	L	L	W	A	I	V	T	V	F	Q	I	L	T	Q	E	D	W	N	K	927
2832	GTC	CTC	TAC	AAT	GGT	ATG	GCC	TCC	ACG	TCG	TCC	TGG	GCG	GCC	CTT	TAT	TTC	ATT	GCC	CTC	2891
928	V	L	Y	N	G	M	A	S	T	S	S	W	A	A	L	Y	F	I	A	L	947
2892	ATG	ACC	TTC	GGC	AAC	TAC	GTG	CTC	TTC	AAT	TTG	CTG	GTC	GCC	ATT	CTG	GTG	GAG	GGC	TTC	2951

948 M T F G N Y V L F N L L V A I L V E G F 967

2952 CAG GCG GAG GAA ATC AGC AAA CGG GAA GAT GCG AGT GGA CAG TTA AGC TGT ATT CAG CTG 3011  
968 Q A E E I S K R E D A S G Q L S C I Q L 987

3012 CCT GTC GAC TCC CAG GGG GGA GAT GCC AAC AAG TCC GAA TCA GAG CCC GAT TTC TTC TCA 3071  
988 P V D S Q G G D A N K S E S E P D F F S 1007

3072 CCC AGC CTG GAT GGT GAT GGG GAC AGG AAG AAG TGC TTG GCC TTG GTG TCC CTG GGA GAG 3131  
1008 P S L D G D G D R K K C L A L V S L G E 1027

3132 CAC CCG GAG CTG CGG AAG AGC CTG CTG CCG CCT CTC ATC ATC CAC ACG GCC GCC ACA CCC 3191  
1028 H P E L R K S L L P P L I I H T A A T P 1047

3192 ATG TCG CTG CCC AAG AGC ACC AGC ACG GGC CTG GGC GAG GCG CTG GGC CCT GCG TCG CGC 3251  
1048 M S L P K S T S T G L G E A L G P A S R 1067

3252 CGC ACC AGC AGC AGC GGG TCG GCA GAG CCT GGG GCG GCC CAC GAG ATG AAG TCA CCG CCC 3311  
1068 R T S S S G S A E P G A A H E M K S P P 1087

3312 AGC GCC CGC AGC TCT CCG CAC AGC CCC TGG AGC GCT GCA AGC AGC TGG ACC AGC AGG CGC 3371  
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3372 TCC AGC CGG AAC AGC CTC GGC CGT GCA CCC AGC CTG AAG CGG AGA AGC CCA AGT GGA GAG 3431  
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3432 CGG CGG TCC CTG TTG TCG GGA GAA GGC CAG GAG AGC CAG GAT GAA GAG GAG AGC TCA GAA 3491  
1128 R R S L L S G E G Q E S Q D E E E S S E 1147

3492 GAG GAG CGG GCC AGC CCT GCG GGC AGT GAC CAT CGC CAC AGG GGG TCC CTG GAG CGG GAG 3551  
1148 E E R A S P A G S D H R H R G S L E R E 1167

3552 GCC AAG AGT TCC TTT GAC CTG CCA GAC ACA CTG CAG GTG CCA GGG CTG CAT CGC ACT GCC 3611  
1168 A K S S F D L P D T L Q V P G L H R T A 1187

3612 AGT GGC CGA GGG TCT GCT TCT GAG CAC CAG GAC TGC AAT GGC AAG TCG GCT TCA GGG CGC 3671  
1188 S G R G S A S E H Q D C N G K S A S G R 1207

3672 CTG GCC CGG GCC CTG CGG CCT GAT GAC CCC CCA CTG GAT GGG GAT GAC GCC GAT GAC GAG 3731  
1208 L A R A L R P D D P P L D G D D A D D E 1227

3732 GGC AAC CTG AGC AAA GGG GAA CGG GTC CGC GCG TGG ATC CGA GCC CGA CTC CCT GCC TGC 3791  
1228 G N L S K G E R V R A W I R A R L P A C 1247

3792 TAC CTC GAG CGA GAC TCC TGG TCA GCC TAC ATC TTC CCT CCT CAG TCC AGG TTC CGC CTC 3851  
1248 Y L E R D S W S A Y I F P P Q S R F R L 1267

3852 CTG TGT CAC CGG ATC ATC ACC CAC AAG ATG TTC GAC CAC GTG GTC CTT GTC ATC ATC TTC 3911  
1268 L C H R I I T H K M F D H V V L V I I F 1287

3912 CTT AAC TGC ATC ACC ATC GCC ATG GAG CGC CCC AAA ATT GAC CCC CAC AGC GCT GAA CGC 3971  
1288 L N C I T I A M E R P K I D P H S A E R 1307

3972 ATC TTC CTG ACC CTC TCC AAT TAC ATC TTC ACC GCA GTC TTT CTG GCT GAA ATG ACA GTG 4031  
1308 I F L T L S N Y I F T A V F L A E M T V 1327

4032 AAG GTG GTG GCA CTG GGC TGG TGC TTC GGG GAG CAG GCG TAC CTG CGG AGC AGT TGG AAC 4091  
1328 K V V A L G W C F G E Q A Y L R S S W N 1347

4092 GTG CTG GAC GGG CTG TTG GTG CTC ATC TCC GTC ATC GAC ATT CTG GTG TCC ATG GTC TCT 4151  
1348 V L D G L L V L I S V I D I L V S M V S 1367

4152 GAC AGC GGC ACC AAG ATC CTG GGC ATG CTG AGG GTG CTG CGG CTG CTG CGG ACC CTG CGC 4211  
1368 D S G T K I L G M L R V L R L L R T L R 1387

4212 CCG CTC AGG GTG ATC AGC CGG GCG CAG GGG CTG AAG CTG GTG GTG GAG ACG CTG ATG TCC 4271  
1388 P L R V I S R A Q G L K L V V E T L M S 1407

4272 TCA CTG AAA CCC ATC GGC AAC ATT GTA GTC ATC TGC TGT GCC TTC TTC ATC ATT TTC GGC 4331  
1408 S L K P I G N I V V I C C A F F I I F G 1427

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1428 I L G V Q L F K G K F F V C Q G E D T R 1447

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2008	P	P	G	R	S	P	L	A	Q	R	P	L	R	R	Q	A	A	I	R	T	2027
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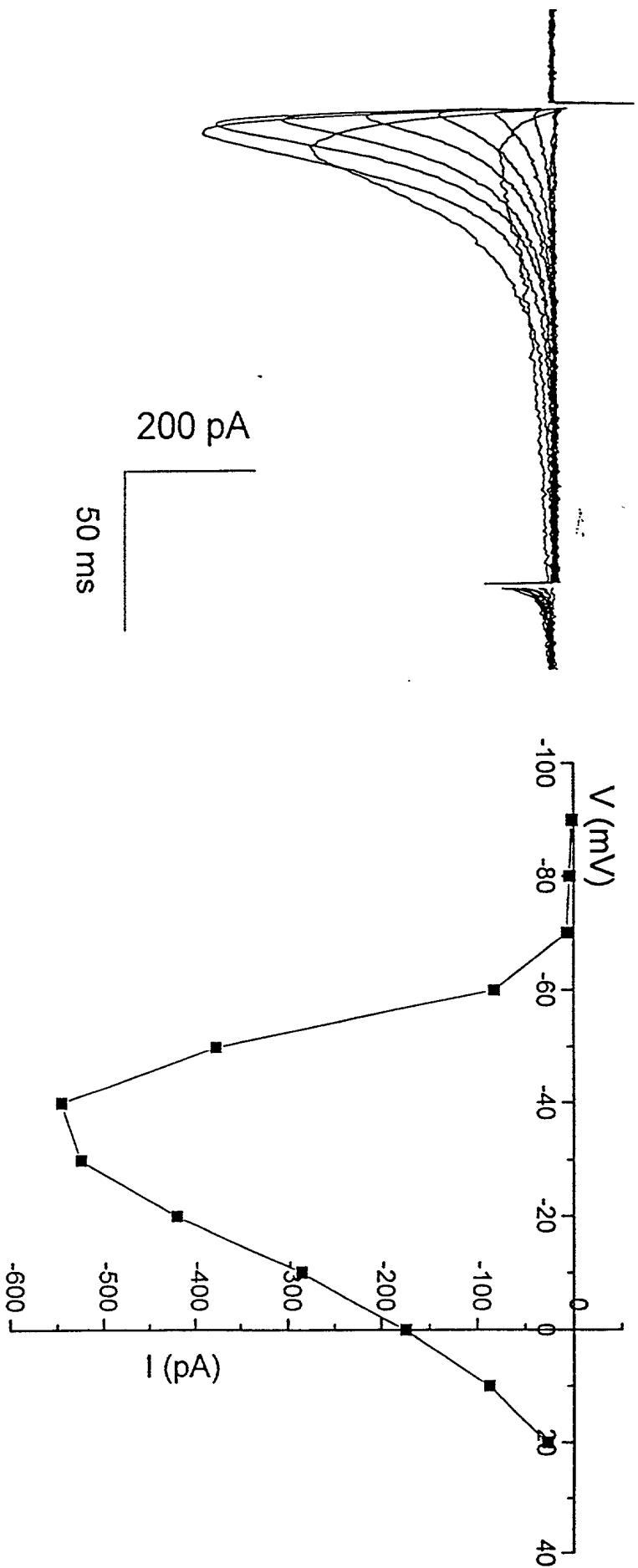


Figure 7  
096410007-070600  
~~Figure 7~~



# COMPARISON OF P-REGIONS

I	II	III	IV	
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LAASQ E GWVYV	QIITQ E GWTDF	ETLSY K GWNVV	RSVTG E DWNDI	NIC-2 (C27F2.3)
EASSQ E GWVFL	QIITQ E GWVDV	EVLSL K GWVEV	RIVTG E DWNKI	Rat-NIC
QCITM E GWTDF	QIITG E DWNSV	TVSTF E GWPEL	RCATG E AWQDI	L-Type Ca Channel
QVITL E GWVDI	QIITQ E DWNKV	VLASK D GWVDI	RVSTG D NWNGI	T-Type Ca Channel
RLMTQ D FWENL	RVLCG E WIETM	QVATF K GWMDI	QITTS A GWDGL	Na Channels

Fig. 8

# SEQUENCE LISTING

<110> Snutch, Terry P.  
Baillie, David L.

<120> NOVEL HUMAN CALCIUM CHANNELS AND RELATED PROBES, CELL  
LINES AND METHODS

<130> NMED.P-001-2(CIP)

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 Cys Pro Cys Cys Gln His Glu Asp Gly Arg Arg Pro Ser Gly Leu Gly  
    725                                      730                                      735  
 Ser Thr Asp Ser Gly Gln Glu Gly Ser Gly Ser Gly Ser Ser Ala Gly  
    740                                      745                                      750  
 Gly Glu Asp Glu Ala Asp Gly Asp Gly Ala Arg Ser Ser Glu Asp Gly  
    755                                      760                                      765  
 Ala Ser Ser Glu Leu Gly Lys Glu Glu Glu Glu Glu Glu Gln Ala Asp  
    770                                      775                                      780  
 Gly Ala Val Trp Leu Cys Gly Asp Val Trp Arg Glu Thr Arg Ala Lys  
 785                                      790                                      795                                      800  
 Leu Arg Gly Ile Val Asp Ser Lys Tyr Phe Asn Arg Gly Ile Met Met

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Ala	Ile	Leu	Val	Asn	Thr	Val	Ser	Met	Gly	Ile	Glu	His	His	Glu	Gln
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Gln	Ala	Asp	Gly	Gly	Leu	Ser	Val	Leu	Arg	Thr	Phe	Arg	Leu	Leu	Arg
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Val	Leu	Lys	Leu	Val	Arg	Phe	Met	Pro	Ala	Leu	Arg	Arg	Gln	Leu	Val
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980				985				990							
Met	Leu	Phe	Ile	Phe	Ile	Phe	Ser	Ile	Leu	Gly	Met	His	Ile	Phe	Gly
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Cys	Lys	Phe	Ser	Leu	Arg	Thr	Asp	Thr	Gly	Asp	Thr	Val	Pro	Asp	Arg
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Lys	Asn	Phe	Asp	Ser	Leu	Leu	Trp	Ala	Ile	Val	Thr	Val	Phe	Gln	Ile
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Leu	Thr	Gln	Glu	Asp	Trp	Asn	Val	Val	Leu	Tyr	Asn	Gly	Met	Ala	Ser
1045				1050				1055							
Thr	Ser	Pro	Trp	Ala	Ser	Leu	Tyr	Phe	Val	Ala	Leu	Met	Thr	Phe	Gly

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Asn Tyr Val Leu Phe Asn Leu Leu Val Ala Ile Leu Val Glu Gly Phe 1075	1080	1085
Gln Ala Glu Val Thr Val Val Leu Ala Glu Glu Ala Pro Pro Gln Gly 1090	1095	1100
Leu Arg Lys Thr Gly Arg Gly Arg Gly Gly Leu Asp Gly Gly Gly Leu 1105	1110	1115 1120
Gln Phe Lys Leu Leu Ala Gly Asn Leu Ser Leu Lys Glu Gly Val Ala 1125	1130	1135
Asp Glu Val Gly Asp Ala Asn Arg Ser Tyr Ser Asp Glu Asp Gln Ser 1140	1145	1150
Ser Ser Asn Ile Glu Glu Phe Asp Lys Leu Gln Glu Gly Leu Asp Ser 1155	1160	1165
Ser Gly Asp Pro Lys Leu Cys Pro Ile Pro Met Thr Pro Asn Gly His 1170	1175	1180
Leu Asp Pro Ser Leu Pro Leu Gly Gly His Leu Gly Pro Ala Gly Ala 1185	1190	1195 1200
Ala Gly Pro Ala Pro Arg Leu Ser Leu Gln Pro Asp Pro Met Leu Val 1205	1210	1215
Ala Leu Gly Ser Arg Lys Ser Ser Val Met Ser Leu Gly Arg Met Ser 1220	1225	1230
Tyr Asp Gln Arg Ser Leu Val Gly Gly Leu Arg Ala Thr Ala Gly Val 1235	1240	1245
Gln Ala Ala Phe Gly His Leu Val Pro Gln Pro Trp Val Cys Leu Trp 1250	1255	1260
Gly Ala Asp Pro Asn Gly Asn Ser Phe Gln Ser Ser Ser Arg Ser Ser 1265	1270	1275 1280
Tyr Tyr Gly Pro Trp Gly Arg Ser Ala Ala Trp Ala Ser Arg Arg Ser 1285	1290	1295
Ser Trp Asn Ser Leu Lys His Lys Pro Pro Ser Ala Glu His Glu Ser 1300	1305	1310
Leu Leu Ser Ala Glu Arg Gly Gly Gly Ala Arg Val Cys Glu Val Ala		

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Ala Asp Glu Gly Pro Pro Arg Ala Ala Pro Leu His Thr Pro His Ala		
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His His Val His His Gly Pro His Leu Ala His Arg His Arg His His		
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Arg Arg Thr Leu Ser Leu Asp Asn Arg Asp Ser Val Asp Leu Ala Glu		
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Leu Val Pro Ala Val Gly Ala His Pro Arg Ala Ala Trp Arg Ala Ala		
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Gly Pro Ala Pro Gly His Glu Asp Cys Asn Gly Arg Met Pro Ser Ile		
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Ala Lys Asp Val Phe Thr Lys Met Gly Asp Arg Gly Asp Arg Gly Glu		
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Asp Glu Glu Glu Ile Asp Tyr Val Ser Gly Gly Gly Ala Glu Gly Asp		
1425	1430	1435 1440
Leu Thr Leu Cys Phe Arg Val Arg Lys Met Ile Asp Val Tyr Lys Pro		
1445	1450	1455
Asp Trp Cys Glu Val Arg Glu Asp Trp Ser Val Tyr Leu Phe Ser Pro		
1460	1465	1470
Glu Asn Arg Leu Arg Asp Leu Gly Trp Val Ser Leu Glu Cys Gln Gly		
1475	1480	1485
Lys Val Gly Asp Leu Val Val Trp Val Tyr Gly Gln Arg Arg Gln Arg		
1490	1495	1500
Gln Thr Ile Ile Ala His Lys Leu Phe Asp Tyr Val Val Leu Ala Phe		
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Ile Phe Leu Asn Cys Ile Thr Ile Ala Leu Glu Arg Pro Gln Ile Glu		
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Ala Gly Ser Thr Glu Arg Ile Phe Leu Thr Val Ser Asn Tyr Ile Phe		
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Thr Ala Ile Phe Val Gly Glu Met Thr Leu Lys Val Val Ser Leu Gly		
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Leu Tyr Phe Gly Glu Gln Ala Tyr Leu Arg Ser Ser Trp Asn Val Leu		

1570	1575	1580
Asp Gly Phe Leu Val Phe Val Ser Ile Ile Asp Ile Val Val Ser Leu		
1585	1590	1595 1600
Ala Ser Ala Gly Gly Ala Lys Ile Leu Gly Val Leu Arg Val Leu Arg		
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Leu Leu Arg Thr Leu Arg Pro Leu Arg Val Ile Ser Arg Ala Pro Gly		
1620	1625	1630
Leu Lys Leu Val Val Glu Thr Leu Ile Ser Ser Leu Lys Pro Ile Gly		
1635	1640	1645
Asn Ile Val Leu Ile Cys Cys Ala Phe Phe Ile Ile Phe Gly Ile Leu		
1650	1655	1660
Gly Val Gln Leu Phe Lys Gly Lys Phe Tyr His Cys Leu Gly Val Asp		
1665	1670	1675 1680
Thr Arg Asn Ile Thr Asn Arg Ser Asp Cys Met Ala Ala Asn Tyr Arg		
1685	1690	1695
Trp Val His His Lys Tyr Asn Phe Asp Asn Leu Gly Gln Ala Leu Met		
1700	1705	1710
Ser Leu Phe Val Leu Ala Ser Lys Asp Gly Trp Val Asn Ile Met Tyr		
1715	1720	1725
Asn Gly Leu Asp Ala Val Ala Val Asp Gln Gln Pro Val Thr Asn His		
1730	1735	1740
Asn Pro Trp Met Leu Leu Tyr Phe Ile Ser Phe Leu Leu Ile Val Ser		
1745	1750	1755 1760
Phe Phe Val Leu Asn Met Phe Val Gly Val Val Val Glu Asn Phe His		
1765	1770	1775
Lys Cys Arg Gln His Gln Glu Ala Glu Glu Ala Arg Arg Arg Glu Glu		
1780	1785	1790
Lys Arg Leu Arg Arg Leu Glu Lys Lys Arg Arg Lys Ala Gln Arg Leu		
1795	1800	1805
Pro Tyr Tyr Ala Thr Tyr Cys His Thr Arg Leu Leu Ile His Ser Met		
1810	1815	1820
Cys Thr Ser His Tyr Leu Asp Ile Phe Ile Thr Phe Ile Ile Cys Leu		

1825 1830 1835 1840

Asn Val Val Thr Met Ser Leu Glu His Tyr Asn Gln Pro Thr  
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<223> human alpha-I partial sequence

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ccatactacc agccggagga ggatgatgag atgcccttca tctgctccct gtcgggcgac 240  
aatgggataa tgggctgcca tgagatcccc ccgctcaagg agcagggccg tgagtgcctgc 300  
ctgtccaagg acgacgtcta cgactttggg gcggggcgcc aggacctcaa tgccagcggc 360  
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aagggtgcca tcagctttga caacatcggt tatgcttgga ttgtcatctt ccaggtgatc 480  
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<211> 189  
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<213> HUMAN

<220>  
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20 25 30  
Gly Val Gln Leu Trp Ala Gly Leu Leu Arg Asn Arg Cys Phe Leu Glu  
35 40 45  
Glu Asn Phe Thr Ile Gln Gly Asp Val Ala Leu Pro Pro Tyr Tyr Gln  
50 55 60  
Pro Glu Glu Asp Asp Glu Met Pro Phe Ile Cys Ser Leu Ser Gly Asp



65	70	75	80
Asn Gly Ile Met Gly Cys His Glu Ile Pro Pro Leu Lys Glu Gln Gly			
85	90	95	
Arg Glu Cys Cys Leu Ser Lys Asp Asp Val Tyr Asp Phe Gly Ala Gly			
100	105	110	
Arg Gln Asp Leu Asn Ala Ser Gly Leu Cys Val Asn Trp Asn Arg Tyr			
115	120	125	
Tyr Asn Val Cys Arg Thr Gly Ser Ala Asn Pro His Lys Gly Ala Ile			
130	135	140	
Ser Phe Asp Asn Ile Gly Tyr Ala Trp Ile Val Ile Phe Gln Val Ile			
145	150	155	160
Thr Leu Glu Gly Trp Val Ala Ile Met Tyr Tyr Val Met Asp Ala Leu			
165	170	175	
Ser Phe Tyr Asn Phe Val Tyr Phe Ile Leu Leu Ile Ile			
180	185		

<210> 21

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<212> DNA

<213> rat

<220>

<223> rat alpha-I partial sequence

<400> 21

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<210> 22

<211> 189

<212> PRT

<213> rat

<220>

<223> rat alpha-I partial sequence

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20 25 30

Gly Val Gln Leu Trp Ala Gly Leu Leu Arg Asn Arg Cys Phe Leu Glu  
35 40 45

Glu Asn Phe Thr Ile Gln Gly Asp Val Ala Leu Pro Pro Tyr Tyr Gln  
50 55 60

Pro Glu Glu Asp Asp Glu Met Pro Phe Ile Cys Ser Leu Thr Gly Asp  
65 70 75 80

Asn Gly Ile Met Gly Cys His Glu Ile Pro Pro Leu Lys Glu Gln Gly  
85 90 95

Arg Glu Cys Cys Leu Ser Lys Asp Asp Val Tyr Asp Phe Gly Ala Gly  
100 105 110

Arg Gln Asp Leu Asn Ala Ser Gly Leu Cys Val Asn Trp Asn Arg Tyr  
115 120 125

Tyr Asn Val Cys Arg Thr Gly Asn Ala Asn Pro His Lys Gly Ala Ile  
130 135 140

Asn Phe Asp Asn Ile Gly Tyr Ala Trp Ile Val Ile Phe Gln Val Ile  
145 150 155 160

Thr Leu Glu Gly Trp Val Glu Ile Met Tyr Tyr Val Met Asp Ala His  
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<212> DNA

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<400> 23



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Arg Gly Glu Gly Gly Gly Gly Pro Pro Cys Ser Leu Asp Tyr Glu Thr  
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 Tyr Asn Ser Ser Ser Asn Thr Thr Cys Val Asn Trp Asn Gln Tyr Tyr  
 340 345 350  
 Thr Asn Cys Ser Ala Gly Glu His Asn Pro Phe Lys Gly Ala Ile Asn  
 355 360 365  
 Phe Asp Asn Ile Gly Tyr Ala Trp Ile Ala Ile Phe Gln Val Ile Thr  
 370 375 380  
 Leu Glu Gly Trp Val Asp Ile Met Tyr Phe Val Met Asp Ala His Ser  
 385 390 395 400  
 Phe Tyr Asn Phe Ile Tyr Phe Ile Leu Leu Ile Ile Val Gly Ser Phe  
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 420 425 430  
 Thr Lys Gln Arg Glu Ser Gln Leu Met Arg Glu Gln Arg Val Arg Phe  
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 Tyr Glu Glu Leu Leu Lys Tyr Leu Val Tyr Ile Leu Arg Lys Ala Ala  
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 Leu Ser Ser Pro Val Ala Arg Ser Gly Gln Glu Pro Gln Pro Ser Gly  
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 His His His His His His His His Tyr His Leu Gly Asn Gly Thr Leu  
 530 535 540  
 Arg Val Pro Arg Ala Ser Pro Glu Ile Gln Asp Arg Asp Ala Asn Gly  
 545 550 555 560  
 Ser Arg Arg Leu Met Leu Pro Pro Pro Ser Thr Pro Thr Pro Ser Gly  
 565 570 575

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Cys	His	Leu	Glu	Pro	Val	Arg	Cys	Gln	Ala	Pro	Pro	Pro	Arg	Cys	Pro	
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Ser	Glu	Ala	Ser	Gly	Arg	Thr	Val	Gly	Ser	Gly	Lys	Val	Tyr	Pro	Thr	
610						615						620				
Val	His	Thr	Ser	Pro	Pro	Pro	Glu	Ile	Leu	Lys	Asp	Lys	Ala	Leu	Val	
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Glu	Val	Ala	Pro	Ser	Pro	Gly	Pro	Pro	Thr	Leu	Thr	Ser	Phe	Asn	Ile	
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Pro	Pro	Gly	Pro	Phe	Ser	Ser	Met	His	Lys	Leu	Leu	Glu	Thr	Gln	Ser	
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Thr	Gly	Ala	Cys	His	Ser	Ser	Cys	Lys	Ile	Ser	Ser	Pro	Cys	Ser	Lys	
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Ala	Asp	Ser	Gly	Ala	Cys	Gly	Pro	Asp	Ser	Cys	Pro	Tyr	Cys	Ala	Arg	
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Asp	Ser	Glu	Ala	Val	Tyr	Glu	Phe	Thr	Gln	Asp	Ala	Gln	His	Ser	Asp	
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Leu	Arg	Asp	Pro	His	Ser	Arg	Arg	Arg	Gln	Arg	Ser	Leu	Gly	Pro	Asp	
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Ala	Glu	Pro	Ser	Ser	Val	Leu	Ala	Phe	Trp	Arg	Leu	Ile	Cys	Asp	Thr	
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Phe	Arg	Lys	Ile	Val	Asp	Ser	Lys	Tyr	Phe	Gly	Arg	Gly	Ile	Met	Ile	
770						775						780				
Ala	Ile	Leu	Val	Asn	Thr	Leu	Ser	Met	Gly	Ile	Glu	Tyr	His	Glu	Gln	
785						790						795			800	
Pro	Glu	Glu	Leu	Thr	Asn	Ala	Leu	Glu	Ile	Ser	Asn	Ile	Val	Phe	Thr	
			805						810						815	
Ser	Leu	Phe	Ala	Leu	Glu	Met	Leu	Leu	Lys	Leu	Leu	Val	Tyr	Gly	Pro	
			820						825						830	



Phe Gly Tyr Ile Lys Asn Pro Tyr Asn Ile Phe Asp Gly Val Ile Val  
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Val Ile Ser Val Trp Glu Ile Val Gly Gln Gln Gly Gly Gly Leu Ser  
850 855 860

Val Leu Arg Thr Phe Arg Leu Met Arg Val Leu Lys Leu Val Arg Phe  
865 870 875 880

Leu Pro Ala Leu Gln Arg Gln Leu Val Val Leu Met Lys Thr Met Asp  
885 890 895

Asn Val Ala Thr Phe Cys Met Leu Leu Met Leu Phe Ile Phe Ile Phe  
900 905 910

Ser Ile Leu Gly Met His Leu Phe Gly Cys Lys Phe Ala Ser Glu Arg  
915 920 925

Asp Gly Asp Thr Leu Pro Asp Arg Lys Asn Phe Asp Ser Leu Leu Trp  
930 935 940

Ala Ile Val Thr Val Phe Gln Ile Leu Thr Gln Glu Asp Trp Asn Lys  
945 950 955 960

Val Leu Tyr Asn Gly Met Ala Ser Thr Ser Ser Trp Ala Ala Leu Tyr  
965 970 975

Phe Ile Ala Leu Met Thr Phe Gly Asn Tyr Val Leu Phe Asn Leu Leu  
980 985 990

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995 1000 1005

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1010 1015 1020

Asp Arg Lys Lys Arg Leu Ala Leu Val Ala Leu Gly Glu His Ala Glu  
1025 1030 1035 1040

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 Cys Phe Leu Pro Glu Asn Phe Ser Leu Pro Leu Ser Val Asp Leu Glu  
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Gln Pro Arg Glu Asn Gly Met Arg Ser Cys Arg Ser Val Pro Thr Leu  
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Tyr Asn Ser Ser Ser Asn Thr Thr Cys Val Asn Trp Asn Gln Tyr Tyr  
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Leu Glu Gly Trp Val Asp Ile Met Tyr Phe Val Met Asp Ala His Ser  
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Cys His Leu Glu Pro Val Arg Cys Gln Ala Pro Pro Pro Arg Ser Pro  
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Glu Val Ala Ala Ser Ser Gly Pro Pro Thr Leu Thr Ser Leu Asn Ile  
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<213> HUMAN

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Ser Pro Ser Glu Ser Pro Ala Ala Glu Arg Gly Ala Glu Leu Gly Ala  
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Asp Glu Glu Gln Arg Val Pro Tyr Pro Ala Leu Ala Ala Thr Val Phe  
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Phe Cys Leu Gly Gln Thr Thr Arg Pro Arg Ser Trp Cys Leu Arg Leu  
85 90 95

Val	Cys	Asn	Pro	Trp	Phe	Glu	His	Val	Ser	Met	Leu	Val	Ile	Met	Leu
			100					105					110		

Asn	Cys	Val	Thr	Leu	Gly	Met	Phe	Arg	Pro	Cys	Glu	Asp	Val	Glu	Cys
		115					120					125			

Gly Ser Glu Arg Cys Asn Ile Leu Glu Ala Phe Asp Ala Phe Ile Phe  
130 135 140

Ala Phe Phe Ala Val Glu Met Val Ile Lys Met Val Ala Leu Gly Leu  
145 150 155 160

Phe Gly Gln Lys Cys Tyr Leu Gly Asp Thr Trp Asn Arg Leu Asp Phe



420				425				430							
Met	Arg	Glu	Gln	Arg	Ala	Arg	His	Leu	Ser	Asn	Asp	Ser	Thr	Leu	Ala
435				440				445							
Ser	Phe	Ser	Glu	Pro	Gly	Ser	Cys	Tyr	Glu	Glu	Leu	Leu	Lys	Tyr	Val
450				455				460							
Gly	His	Ile	Phe	Arg	Lys	Val	Lys	Arg	Arg	Ser	Leu	Arg	Leu	Tyr	Ala
465				470				475				480			
Arg	Trp	Gln	Ser	Arg	Trp	Arg	Lys	Lys	Val	Asp	Pro	Ser	Ala	Val	Gln
485				490				495							
Gly	Gln	Gly	Pro	Gly	His	Arg	Gln	Arg	Arg	Ala	Gly	Arg	His	Thr	Ala
500				505				510							
Ser	Val	His	His	Leu	Val										
515															

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<210> 34
<211> 1080
<212> DNA
<213> HUMAN
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<400> 34						
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acagcctcaa	gcacaagccg	ccgtcggcgg	agcatgagtc	cctgtctctct	gcggagcgcg	180
gcggcgggcg	ccgggtctgc	gaggttgccg	cggacgaggg	gccgccgcgg	gccgcacccc	240
tgcacacccc	acacgcccac	cacattcatc	acggggccca	tctggcgcac	cgccaccgcc	300
accaccgcgg	gacgtgtcc	ctcgacaaca	gggactcggt	ggacctggcc	gagctggtgc	360
ccgcggtggg	cgcccacccc	cgggcgcgct	ggagggcggc	aggcccggcc	ccggggcatg	420
aggactgcaa	tggcaggatg	cccagcatcg	caaagacgt	cttcaccaag	atgggcgacc	480
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agcaggcgta	cctacgcagc	agctggaacg	tgctggatgg	ctttcttgtc	ttcgtgtcca	900
tcacgcacat	cgtggtgtcc	ctggcctcag	ccgggggagc	caagatcttg	ggggtcctcc	960
gagtccttgcg	gctcctgcgc	accctacgcc	ccctgcgtgt	catcagccgg	gcgccggggc	1020
tgaagctggg	ggtggagaca	ctcatctcct	ccctcaagcc	catcggcaac	atcgtgtctca	1080

$$\begin{aligned} \langle 210 \rangle & 35 \\ \langle 211 \rangle & 359 \end{aligned}$$

<212> PRT  
<213> HUMAN

<400> 35

Ser Val Met Ser Leu Gly Arg Met Ser Tyr Asp Gln Arg Ser Leu Ser  
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Ser Ser Arg Ser Ser Tyr Tyr Gly Pro Trp Gly Arg Ser Ala Ala Trp  
20 25 30

Ala Ser Arg Arg Ser Ser Trp Asn Ser Leu Lys His Lys Pro Pro Ser  
35 40 45

Ala Glu His Glu Ser Leu Leu Ser Ala Glu Arg Gly Gly Gly Ala Arg  
50 55 60

Val Cys Glu Val Ala Ala Asp Glu Gly Pro Pro Arg Ala Ala Pro Leu  
65 70 75 80

His Thr Pro His Ala His His Ile His His Gly Pro His Leu Ala His  
85 90 95

Arg His Arg His His Arg Arg Thr Leu Ser Leu Asp Asn Arg Asp Ser  
100 105 110

Val Asp Leu Ala Glu Leu Val Pro Ala Val Gly Ala His Pro Arg Ala  
115 120 125

Ala Trp Arg Ala Ala Gly Pro Ala Pro Gly His Glu Asp Cys Asn Gly  
130 135 140

Arg Met Pro Ser Ile Ala Lys Asp Val Phe Thr Lys Met Gly Asp Arg  
145 150 155 160

Gly Asp Arg Gly Glu Asp Glu Glu Glu Ile Asp Tyr Thr Leu Cys Phe  
165 170 175

Arg Val Arg Lys Met Ile Asp Val Tyr Lys Pro Asp Trp Cys Glu Val  
180 185 190

Arg Glu Asp Trp Ser Val Tyr Leu Phe Ser Pro Glu Asn Arg Phe Arg  
195 200 205

Val Leu Cys Gln Thr Ile Ile Ala His Lys Leu Phe Asp Tyr Val Val  
210 215 220

Leu Ala Phe Ile Phe Leu Asn Cys Ile Thr Ile Ala Leu Glu Arg Pro  
225 230 235 240

Gln Ile Glu Ala Gly Ser Thr Glu Arg Ile Phe Leu Thr Val Ser Asn  
245 250 255

Tyr Ile Phe Thr Ala Ile Phe Val Gly Glu Met Thr Leu Lys Val Val  
260 265 270

Ser Leu Gly Leu Tyr Phe Gly Glu Gln Ala Tyr Leu Arg Ser Ser Trp  
275 280 285

Asn Val Leu Asp Gly Phe Leu Val Phe Val Ser Ile Ile Asp Ile Val  
290 295 300

Val Ser Leu Ala Ser Ala Gly Gly Ala Lys Ile Leu Gly Val Leu Arg  
305 310 315 320

Val Leu Arg Leu Leu Arg Thr Leu Arg Pro Leu Arg Val Ile Ser Arg  
325 330 335

Ala Pro Gly Leu Lys Leu Val Val Glu Thr Leu Ile Ser Ser Leu Lys  
340 345 350

Pro Ile Gly Asn Ile Val Leu  
355